Reference-quality diploid genomes without *de novo* assembly

Michael Schatz

January 16, 2018 PAG Bioinformatics Workshop



@mike_schatz / #PAGXXVI

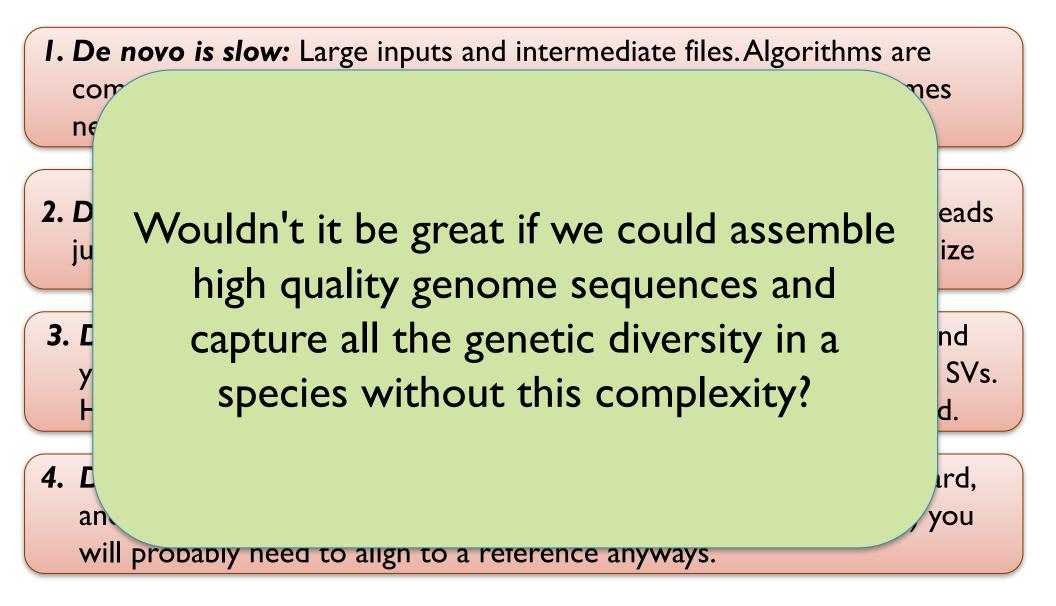


De novo is necessary for the first genome of a species, but is it really necessary for genomes 2 through N?

- **I. De novo is slow:** Large inputs and intermediate files. Algorithms are complex to compare all the reads to each other. Mammalian genomes need thousands of core hours and terabytes of space
- **2.** De novo is demanding: Make the libraries just right, sequence the reads just right, set the parameters just right, launch and relaunch to optimize
- 3. De novo is unpredictable: Add a little more (or a little less) data, and your contig N50 drops in half. Errors creep in ranging from SNPs to SVs. Heuristics break when the data or genome structure are unexpected.
- **4.** De novo is just the beginning. Annotation from scratch is really hard, and to use it (variant analysis/selection analysis/regulatory analysis) you will probably need to align to a reference anyways.



De novo is necessary for the first genome of a species, but is it really necessary for genomes 2 through N?

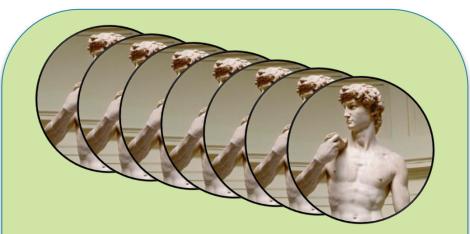


Reference-Guided Assembly



I. High quality reference

- Contig N50 over IMbp
- Scaffold N50 over 10Mbp
- High Quality Gene Annotation (See VGP definition)
- Your sample is sufficiently similar (~99% identity)



2. Sample specific data

- <u>SNPs and Indels</u>: Illumina-based (Illumina PE or IOX)
- <u>Structural Variants</u>: Long Reads (PacBio or ONT)
- <u>Phasing Data</u>: I0X and/or HiC; trios when available

Data requirements similar to de novo, but less demanding, more accurate, and more predictable

Comparative Genome Assembly ("AMOScmp") Pop et al (2004) Briefings in Bioinformatics. Sep;5(3):237-48.

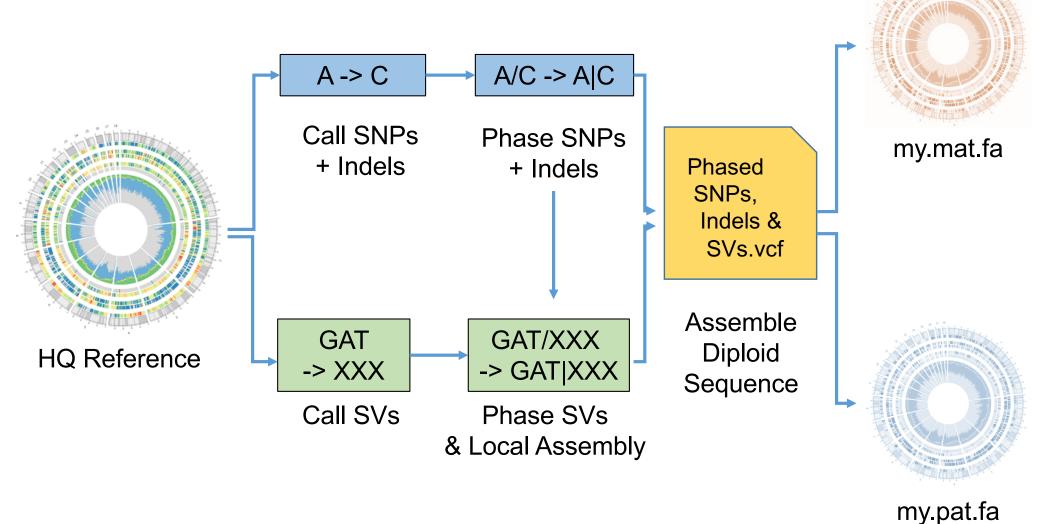






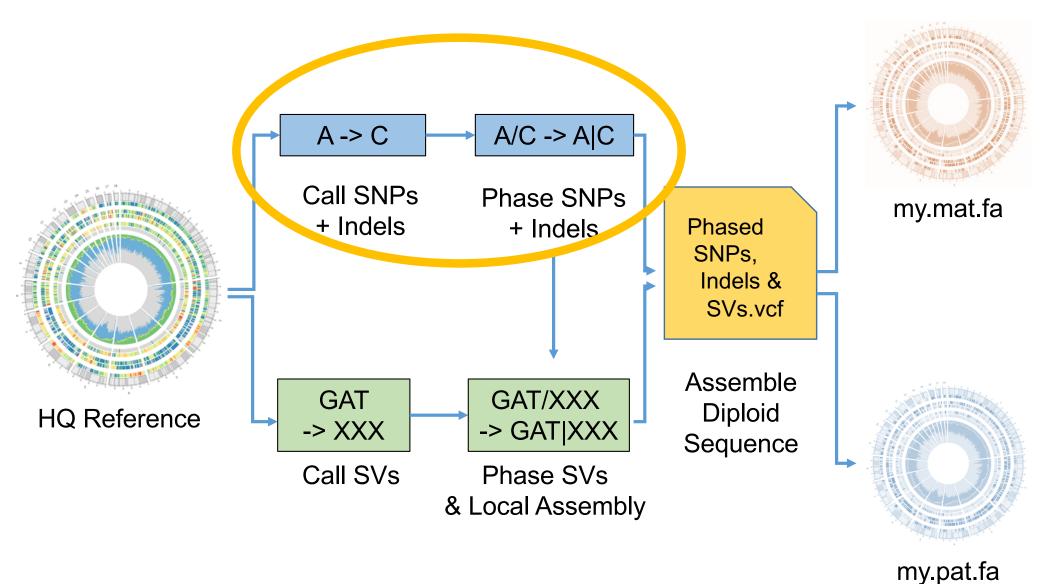




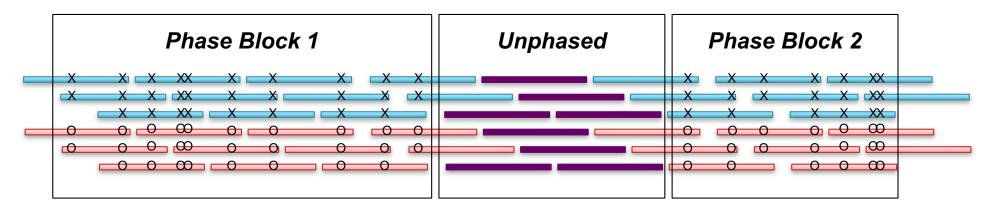




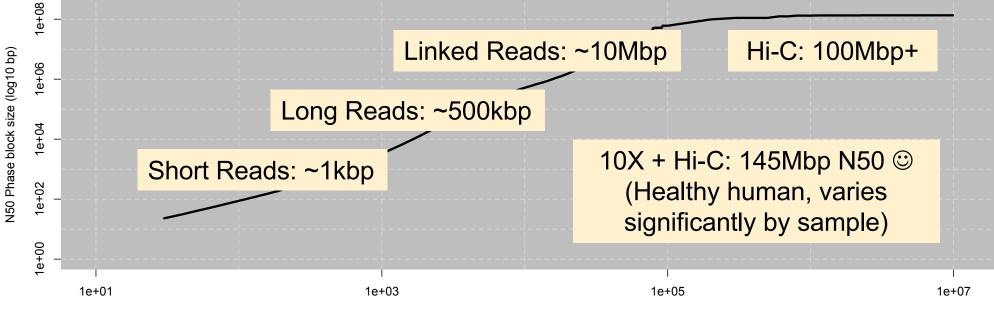




Phasing Results



NA12878 Optimal phase block length increases with read length

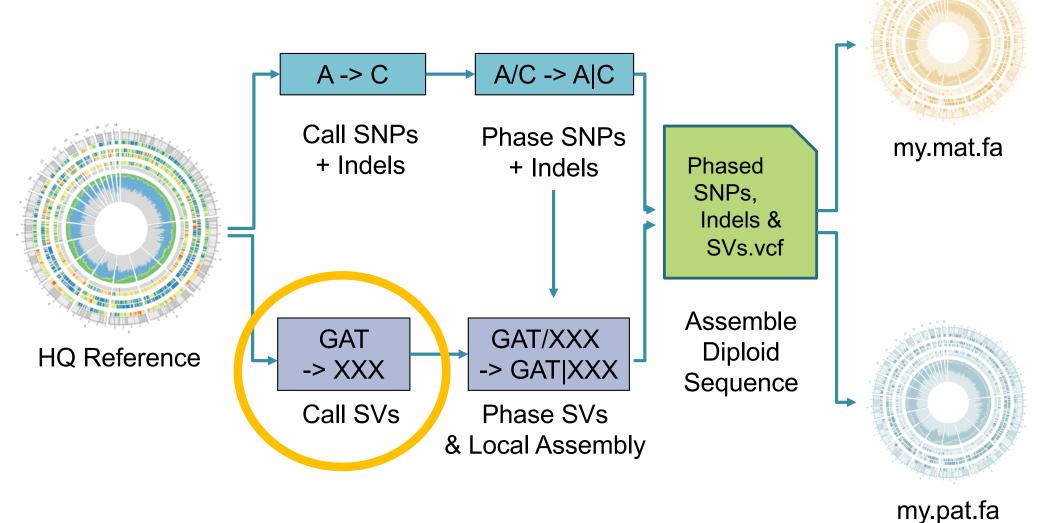


Read length (log10 bp)

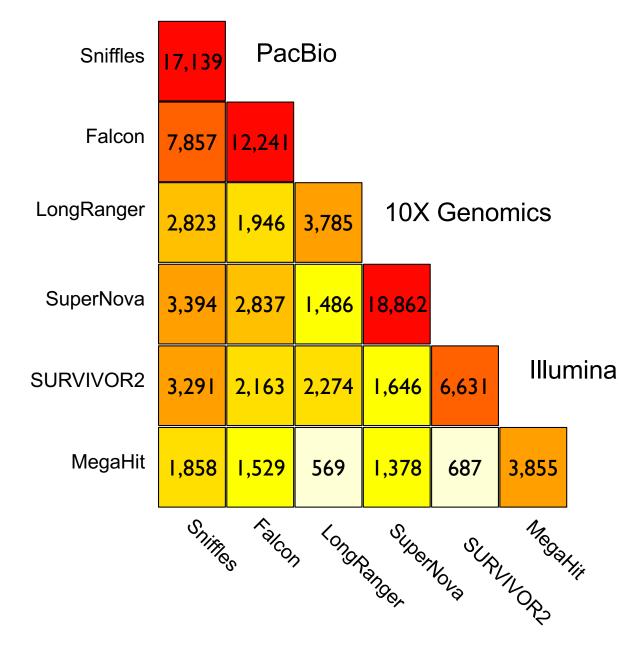
HapCUT2: robust and accurate haplotype assembly for diverse sequencing technologies Edge, P, Bafna, V, Bansal, V (2016) Genome Research. doi: 10.1101/gr.213462.116







SVs using Short, Long and Linked Reads



Main Diagonal

Calls per tool

Outer triplets

• Concordance by Technology

Inner triplets

- Concordance by Assembly
- Concordance by Mappers

Overall:

 Lonnnnnng reads give the most variants with the best concordance ⁽³⁾



NGMLR + Sniffles



BWA-MEM:



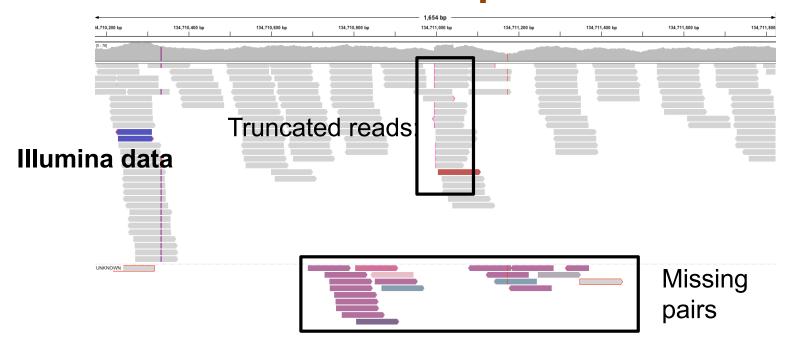
NGMLR:

195,235,300 kp 193,235,400 kp	193,225,530 kp	- 805 bp	193,235,716 bp	
	22	7	II · I I I	
	- 215 - 224 - 224	1		T THE
				-1 1 11-1 1
				· · I · · ·
	22			
	- 22 - 224 - 225 - 21			
	22	7		i da d
	113	1		
	-11 -12 -12			111111-1
				I → I -
		7		1.1.10
	- 128	7		-11 11
	22 221 - 221	1		
		7		
			=	117 -
		26		1
an Tanàn ao Mandri P		1		I-I

NGMLR: Convex gap penalty to balance frequent small sequencing errors with larger SVs Sniffles: Scan within and between split reads to accurately find SVs (Ins, Del, Dup, Inv, Trans) Mendelian concordance >95%, experimental validation also very high

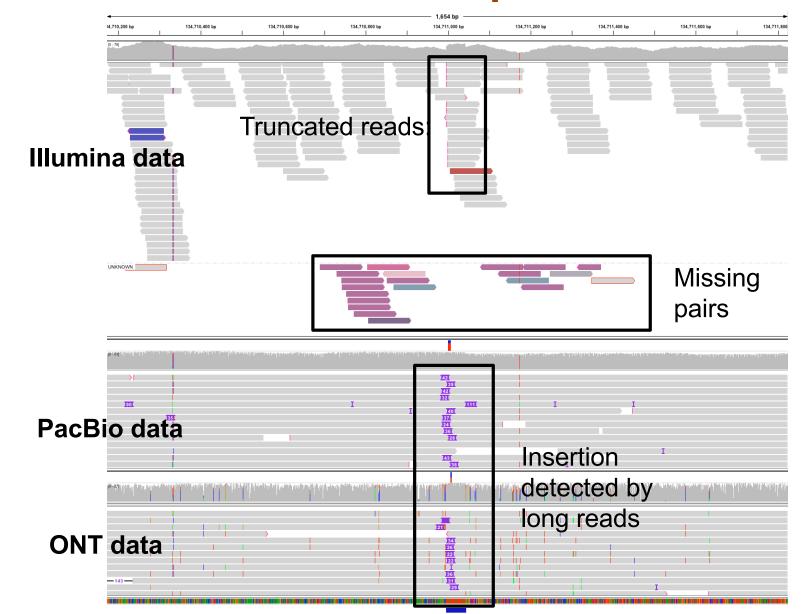
Accurate detection of complex structural variations using single molecule sequencing Sedlazeck, Rescheneder et al (2017) bioRxiv https://doi.org/10.1101/169557

No more false positives!



Accurate detection of complex structural variations using single molecule sequencing Sedlazeck, Rescheneder et al (2017) bioRxiv https://doi.org/10.1101/169557

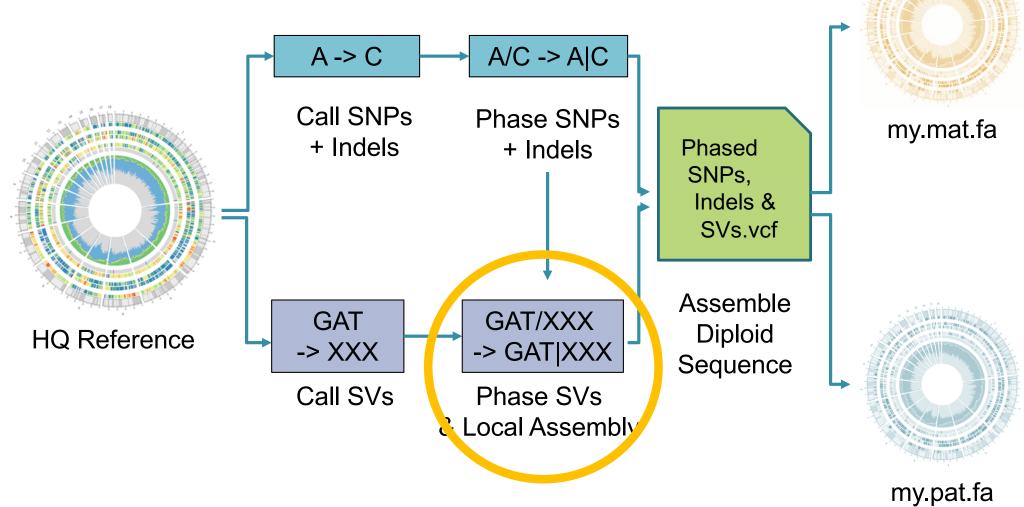
No more false positives!



Accurate detection of complex structural variations using single molecule sequencing Sedlazeck, Rescheneder et al (2017) bioRxiv https://doi.org/10.1101/169557

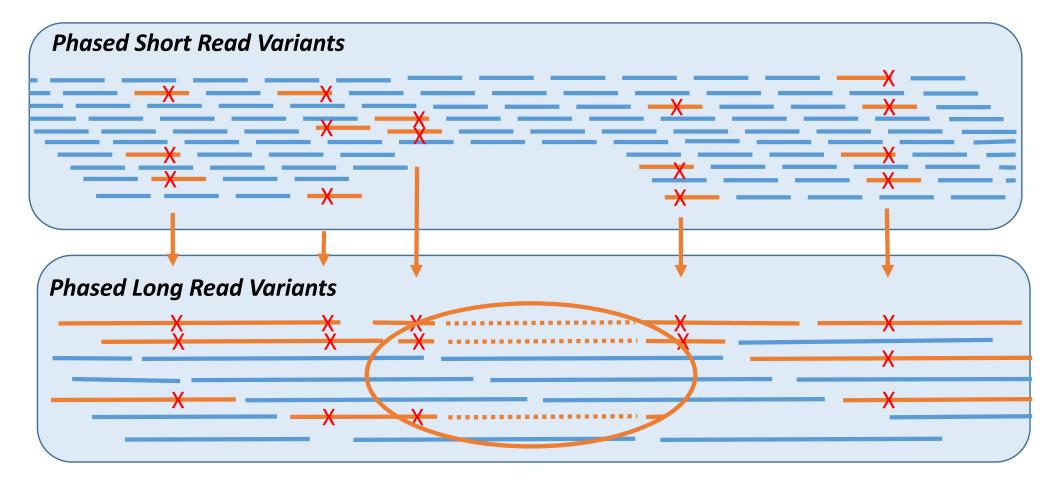






Local Assembly and SV Phasing

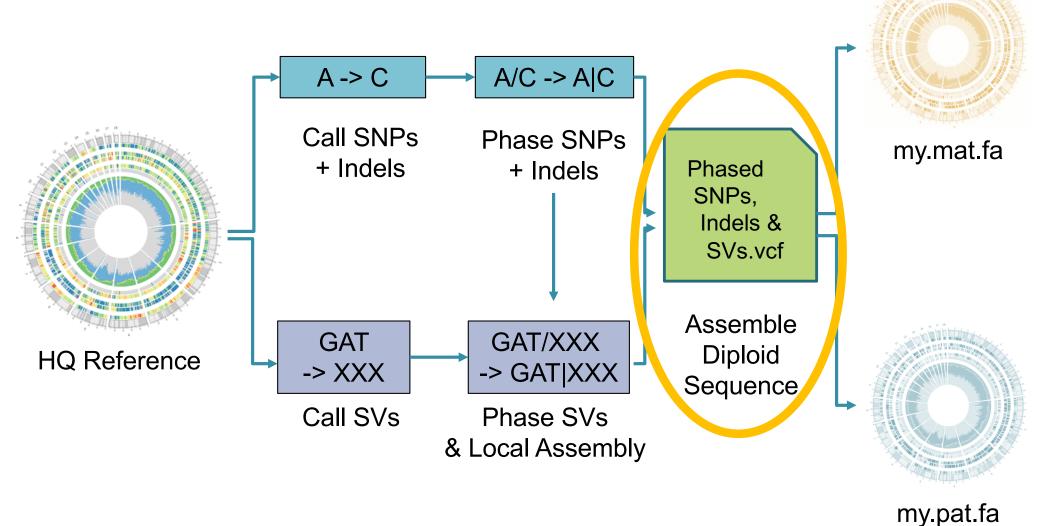
Transfer the phasing of the short read variants to the long reads The phased long reads allow the SVs to be phased



Phase SVs: Make sure SVs are associated with the correct haplotype *Local Assembly*: Refine sequence of insertions, resolve complex nested variants

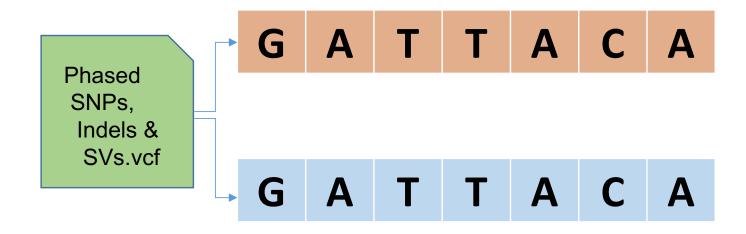






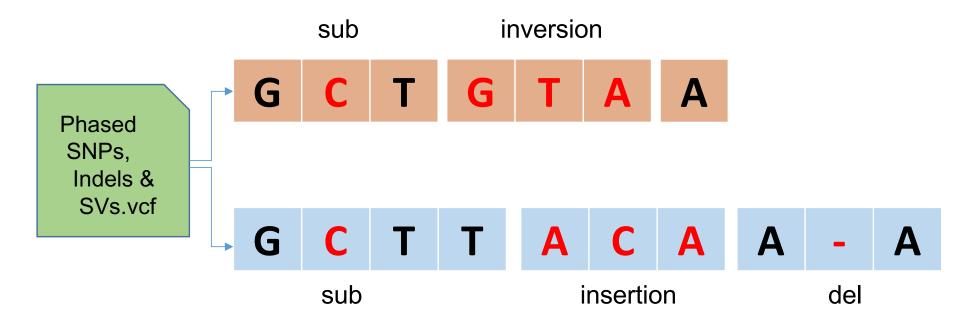
Assembling a "Perfect" Personalized Diploid Genome

Carefully "stitch" the phased variants into the reference genome at the right position to create a pair of phased chromosome fasta files



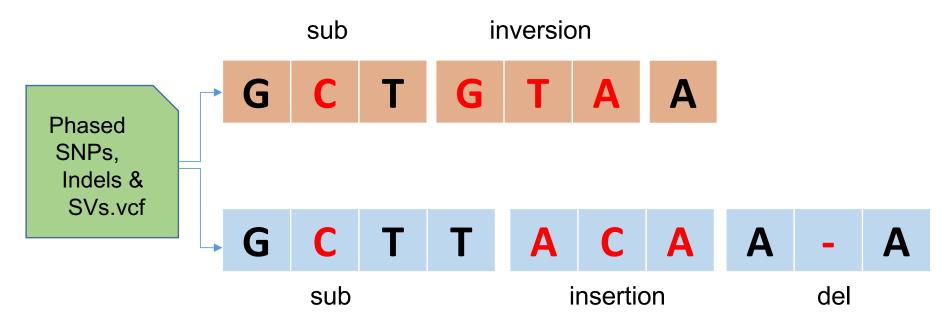
Assembling a "Perfect" Personalized Diploid Genome

Carefully "stitch" the phased variants into the reference genome at the right position to create a pair of phased chromosome fasta files



Assembling a "Perfect" Personalized Diploid Genome

Carefully "stitch" the phased variants into the reference genome at the right position to create a pair of phased chromosome fasta files



Stitching based on AlleleSeq pipeline enhanced for SVs (Rozowsky et al, 2011)

• Maintains a mapping from reference to personal genome coordinates to make lift over of annotation straightforward to compute

Using IOX + HiC + PacBio, assemble essentially perfect diploid human genomes with haplotypes spanning entire chromosomes

 Phased diploid genome can be aligned or aligned against just like a de novo genome assembly

Applications

Expression & Regulation



Foundation for mapping functional data

- Discover novel genes and gene fusions
- Analyze differential expression in CNVs
- Discover new regulatory regions
- Analyze allele-specific expression

Population Genetics



Framework for GWAS of Structural Variations

- Identified SVs in >900 accessions using short reads
- Assembling the top 50 lines using long & linked reads
- Perform GWAS of breeding traits

Polyploidy



Studying heterozygosity in sugarcane

- Have a high quality PacBio-based assembly of POJ2878 using FALCON (140kbp N50)
- Developing new methods for phasing (9-14 copies of each chromosome)

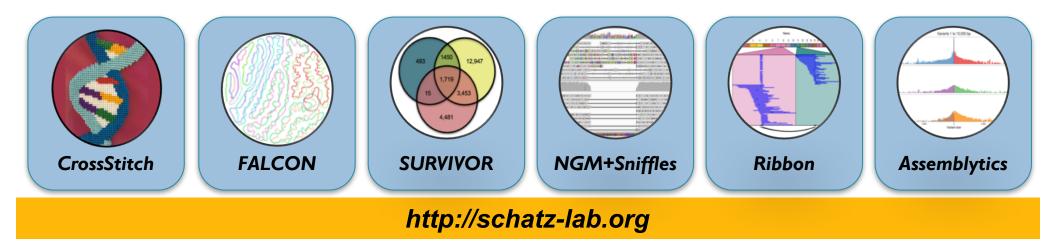
Reference-quality Genomes without de novo assembly

De novo assembly is essential for exploring new species

• Reference-free or mapping to a distant reference is difficult to impossible

But once the first genome of a species has been assembled, shouldn't the second genome be a little easier?

- Use the right combination of data to capture and phase all types of variants
- Overnight analysis to create a high quality personalized genome that are more accurate, more predictable, and easier to use than a de novo assembly
- The personalized diploid genome will be a platform for functional and evolutionary analysis in many species



Acknowledgements

Schatz Lab

Mike Alonge Amelia Bateman Charlotte Darby Han Fang Michael Kirsche Sam Kovaka Laurent Luo Srividya Ramakrishnan T. Rhyker Ranallo-Benavide ***Your Name Here***

Baylor Medicine

Fritz Sedlazeck

University of Vienna

Arndt von Haeseler Philipp Rescheneder

DNAnexus Maria Nattestad

CSHL

Gingeras Lab Jackson Lab Lippman Lab Lyon Lab Martienssen Lab McCombie Lab **Tuveson Lab** Ware Lab Wigler Lab

SBU

Skiena Lab Patro Lab

GRC

Roderic Guido Alessandra Breschi Anna Vlasova

Yale

Gerstein Lab

JHU

Battle Lab Langmead Lab Leek Lab Salzberg Lab Taylor Lab Timp Lab Wheelan Lab

Cornell Susan McCouch Lyza Maron Mark Wright

OICR

John McPherson Karen Ng **Timothy Beck** Yogi Sundaravadanam

PacBio Greg Concepcion





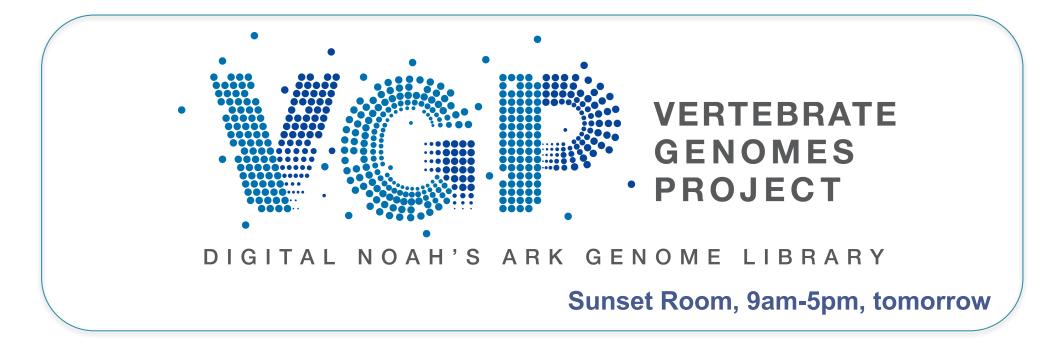
National Human Genome Research Institute



SFARI SIMONS FOUNDATION AUTISM RESEARCH INITIATIVE



ALFRED P. SLOAN FOUNDATION



Thank you! @mike_schatz