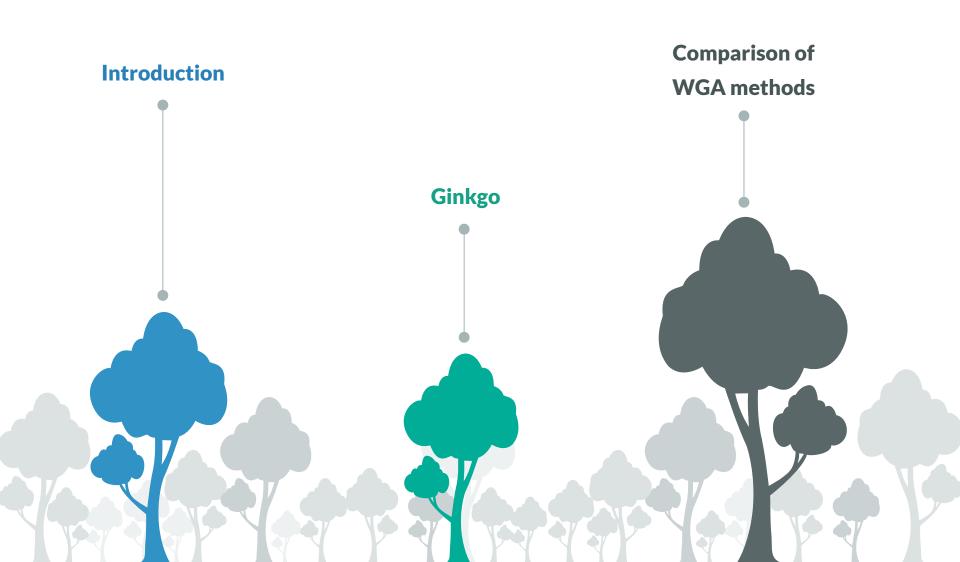
# **Ginkgo**—Interactive analysis and quality assessment of single-cell CNV data



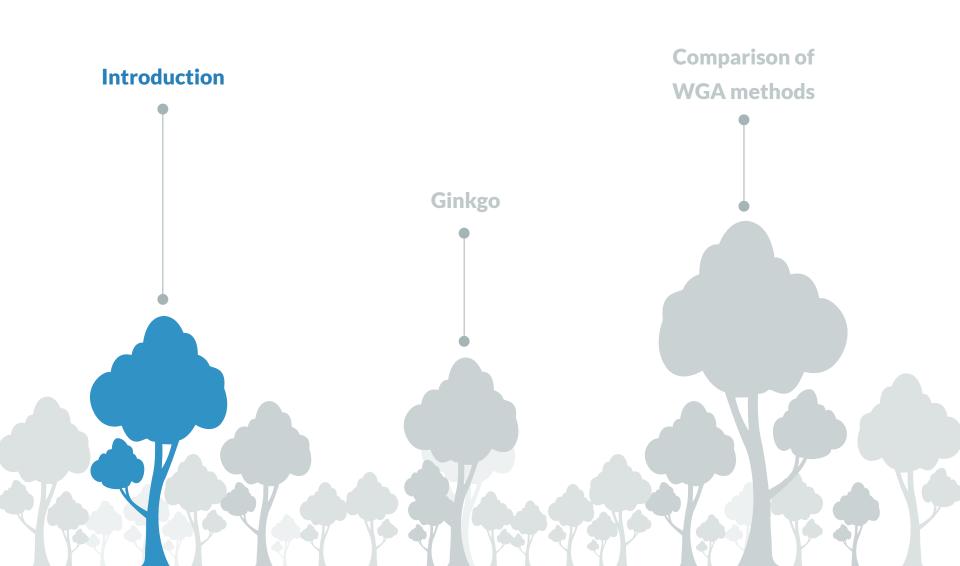
Robert Aboukhalil, Tyler Garvin, Jude Kendall, Timour Baslan, Gurinder S. Atwal, Jim Hicks, Michael Wigler, Michael C. Schatz

CSH Cold Spring Harbor Laboratory

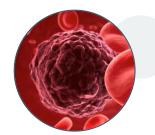
# Outline



# Outline



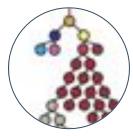
### Single-cell sequencing



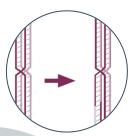
Circulating tumor cells



Neuronal mosaicism

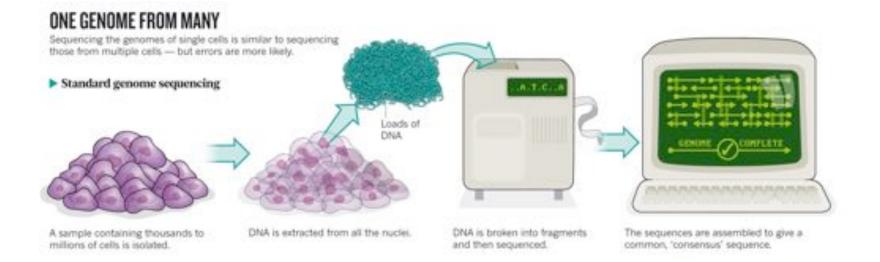


Clonal evolution in tumors

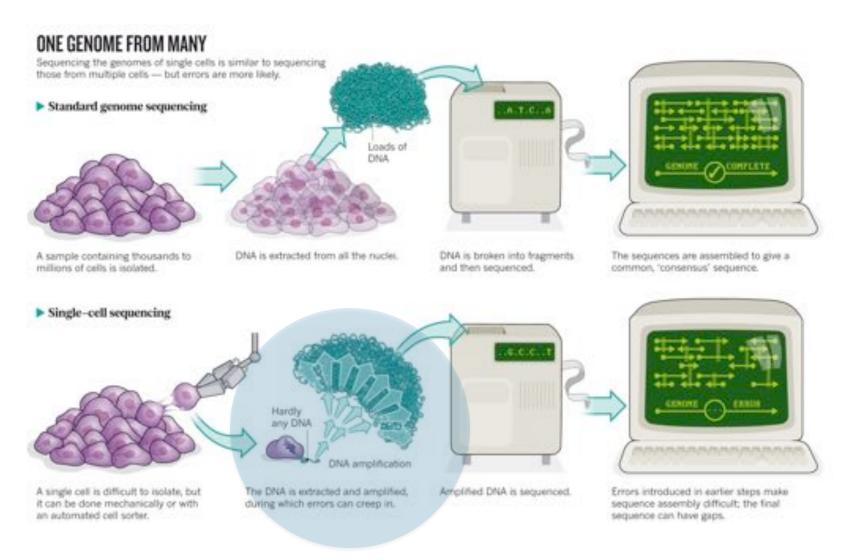


Recombination/ crossover in germ cells

#### Single-cell vs. bulk sequencing



#### Single-cell vs. bulk sequencing





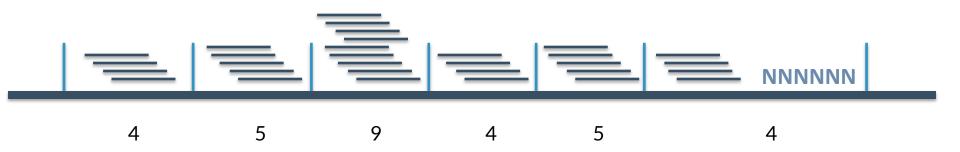
#### Low coverage allows us to study copy-number variants <1X coverage, often <0.1X

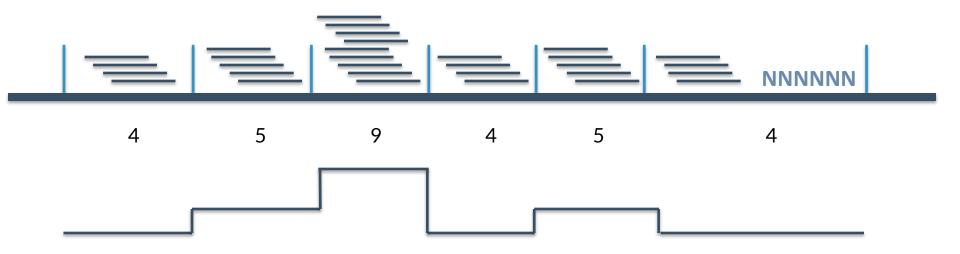
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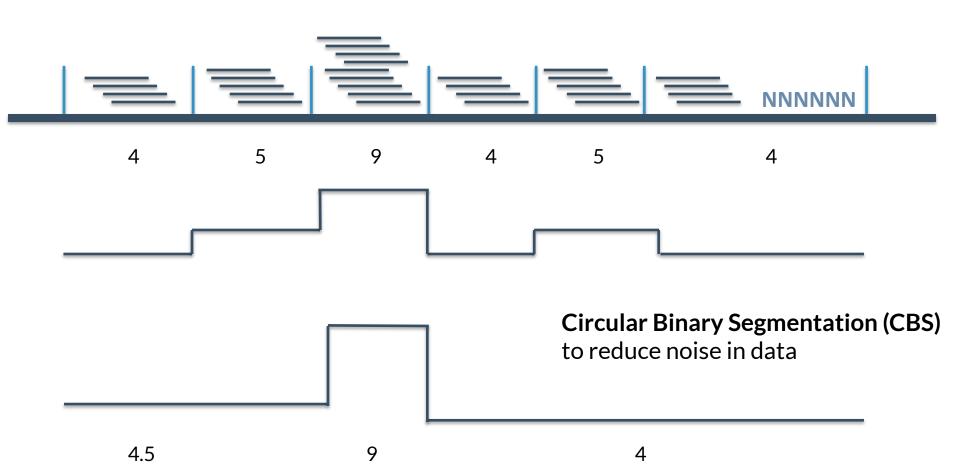
#### Low coverage allows us to study copy-number variants <1X coverage, often <0.1X

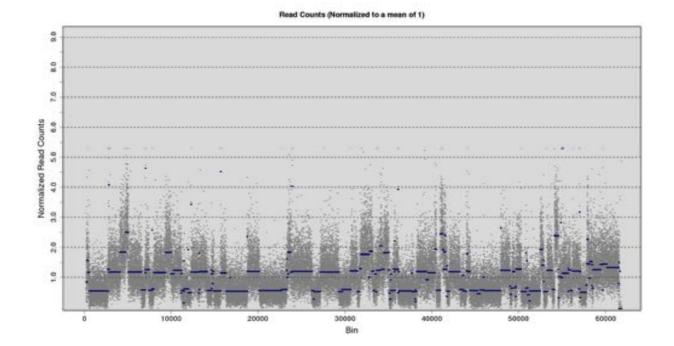


#### Divide genome into "bins" with ~50 - 100 reads / bin



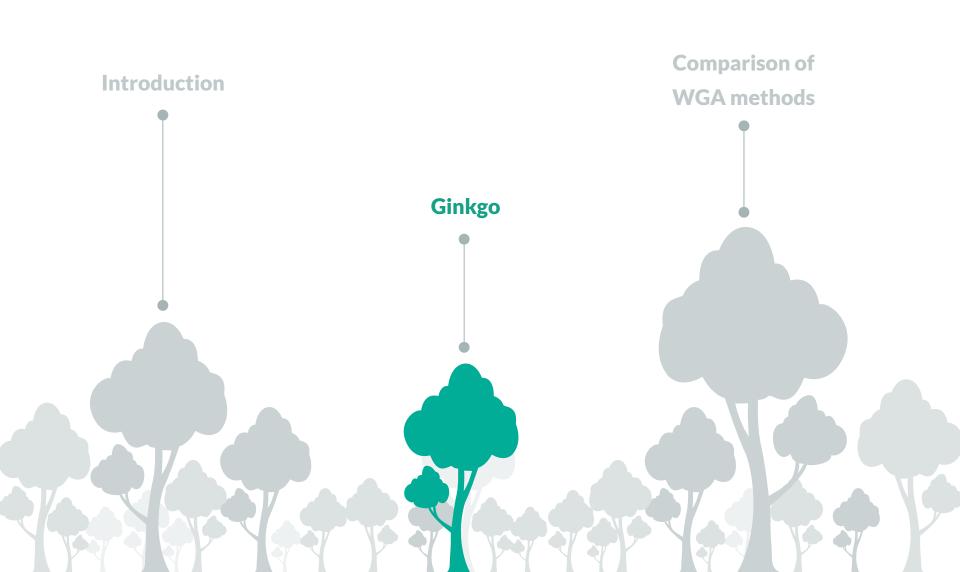


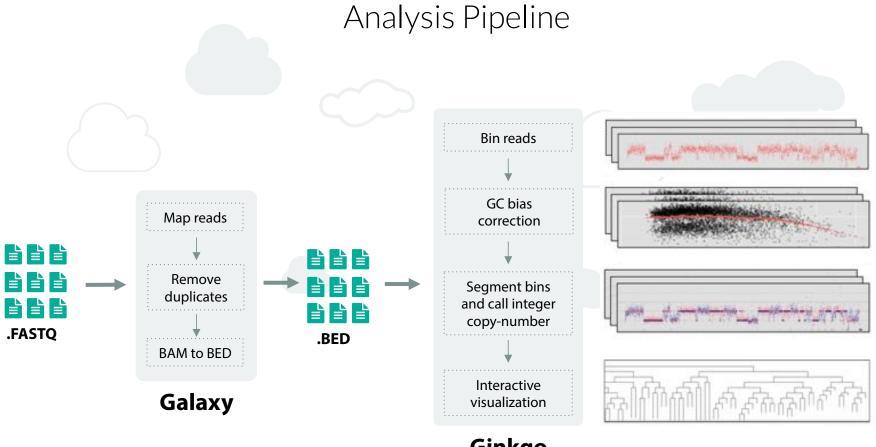




We can estimate integer copy-number states by scaling the profile and minimizing the sum of squares error

# Outline





Ginkgo

## Ginkgo Demo

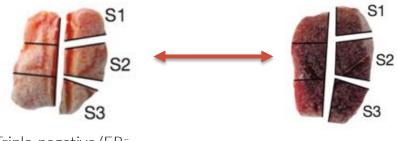
#### Sample dataset



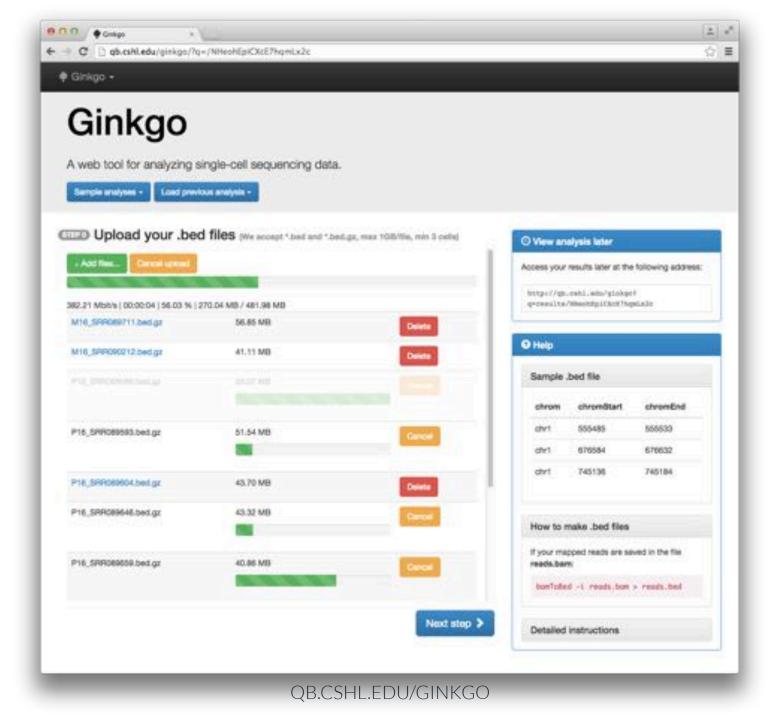
doi:10.1038/nature098.07

#### Tumour evolution inferred by single-cell sequencing

Nicholas Navin<sup>1,2</sup>, Jude Kendall<sup>1</sup>, Jennifer Troge<sup>1</sup>, Peter Andrews<sup>1</sup>, Linda Rodgers<sup>1</sup>, Jeanne McIndoo<sup>1</sup>, Kerry Cook<sup>1</sup>, Asya Stepansky<sup>1</sup>, Dan Levy<sup>1</sup>, Diane Esposito<sup>1</sup>, Lakshmi Muthuswamy<sup>3</sup>, Alex Krasnitz<sup>1</sup>, W. Richard McComble<sup>1</sup>, James Hicks<sup>1</sup> & Michael Wigler<sup>1</sup>



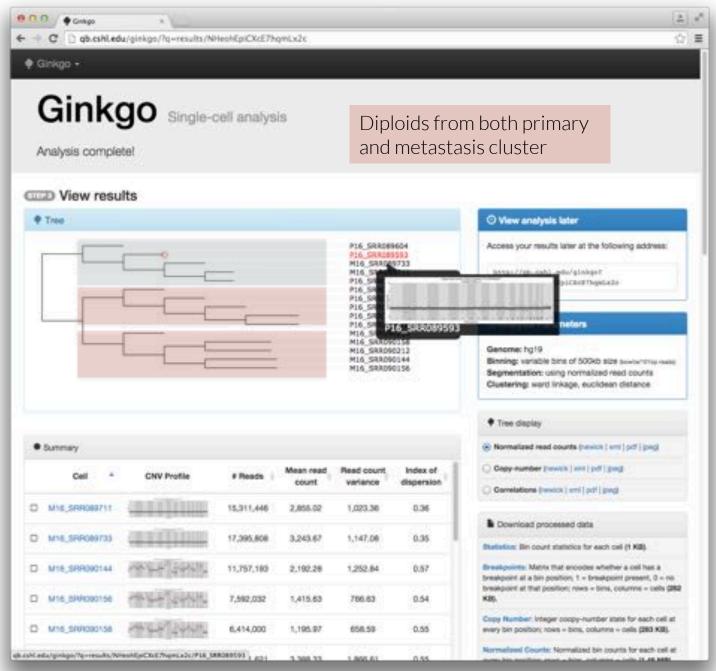
Triple-negative (ER<sup>-</sup>, PR<sup>-</sup>, HER2<sup>-</sup>) ductal carcinoma

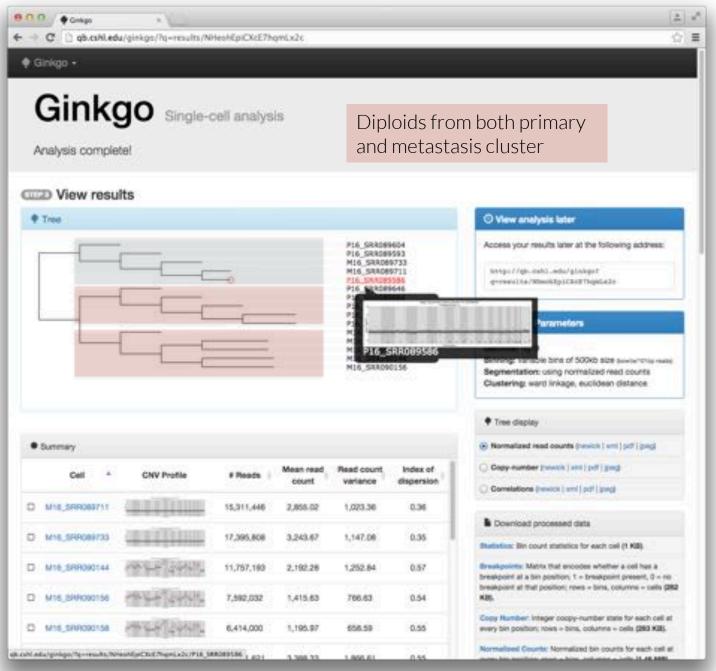


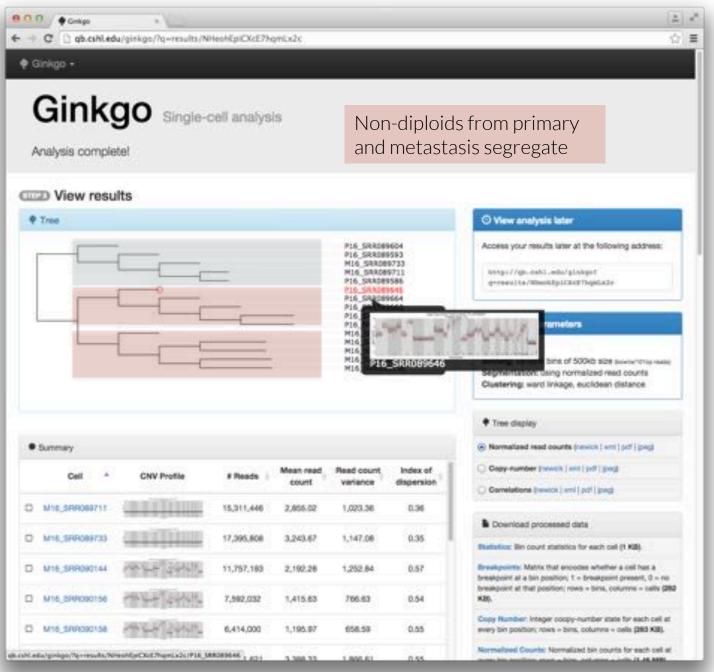
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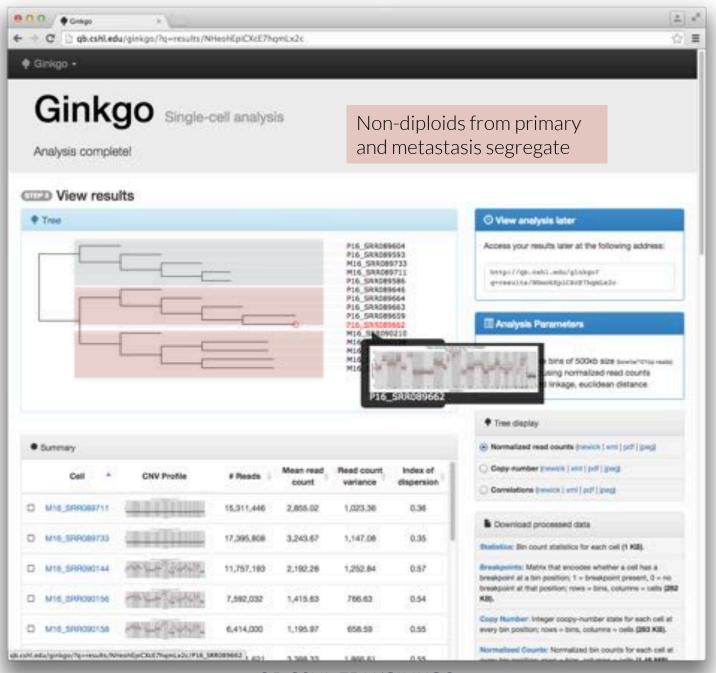
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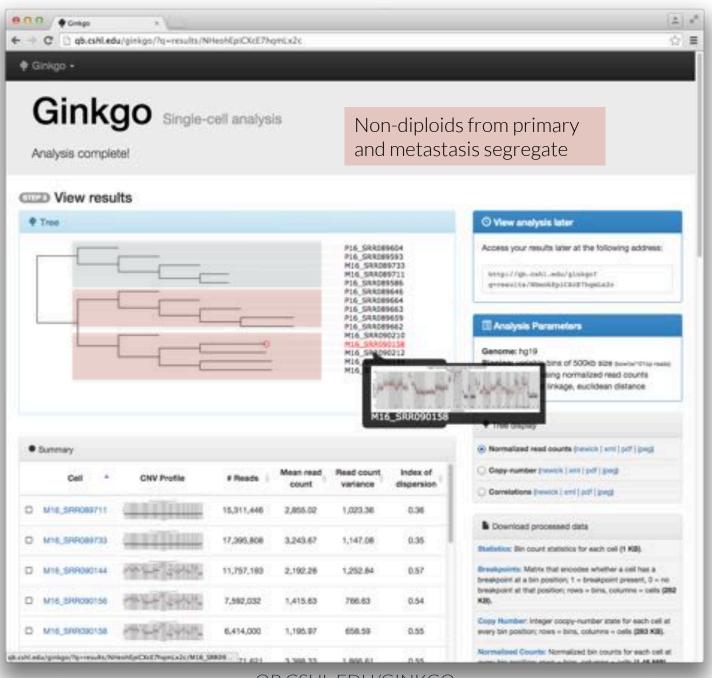
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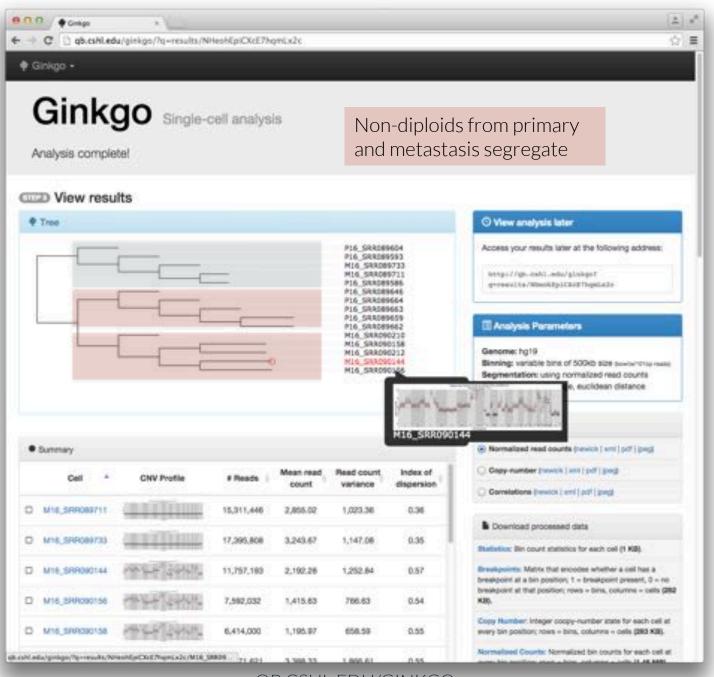


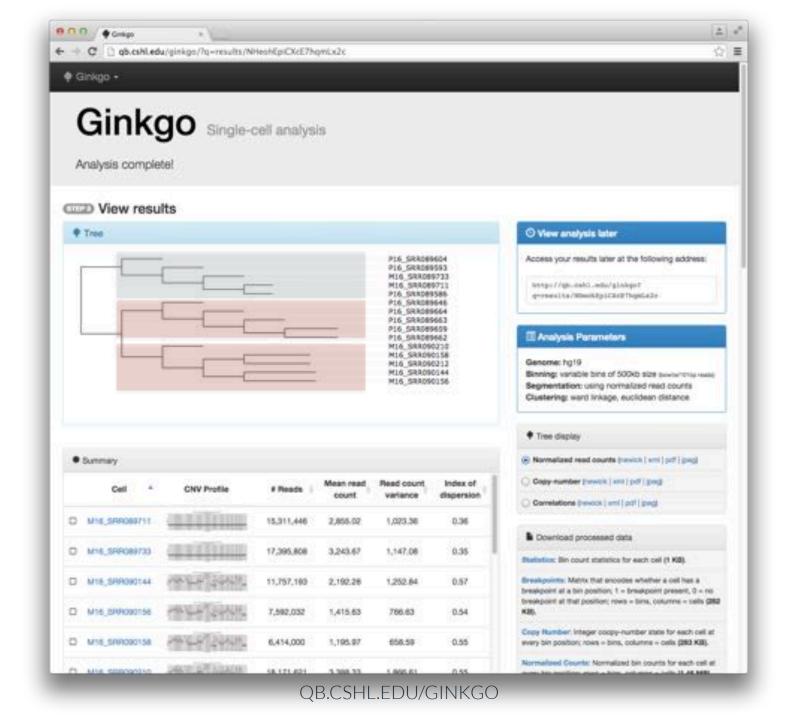










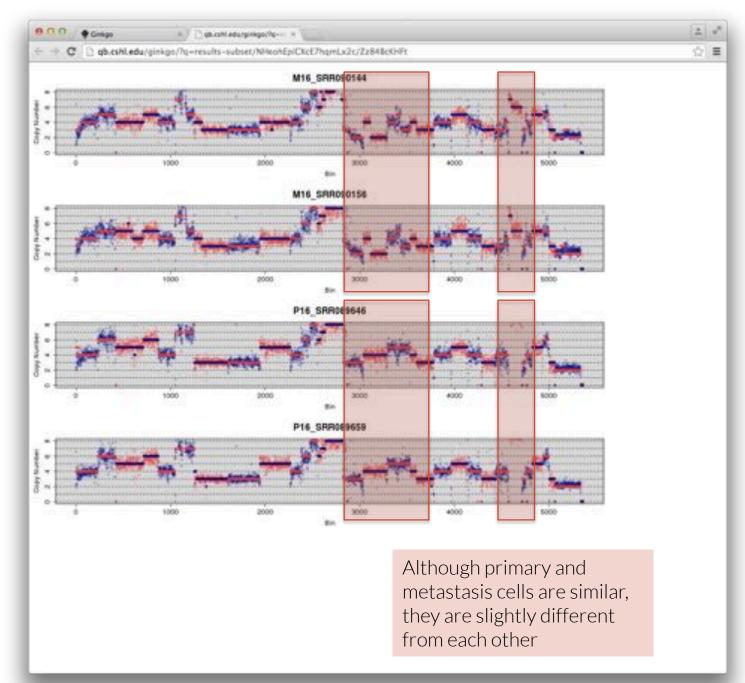


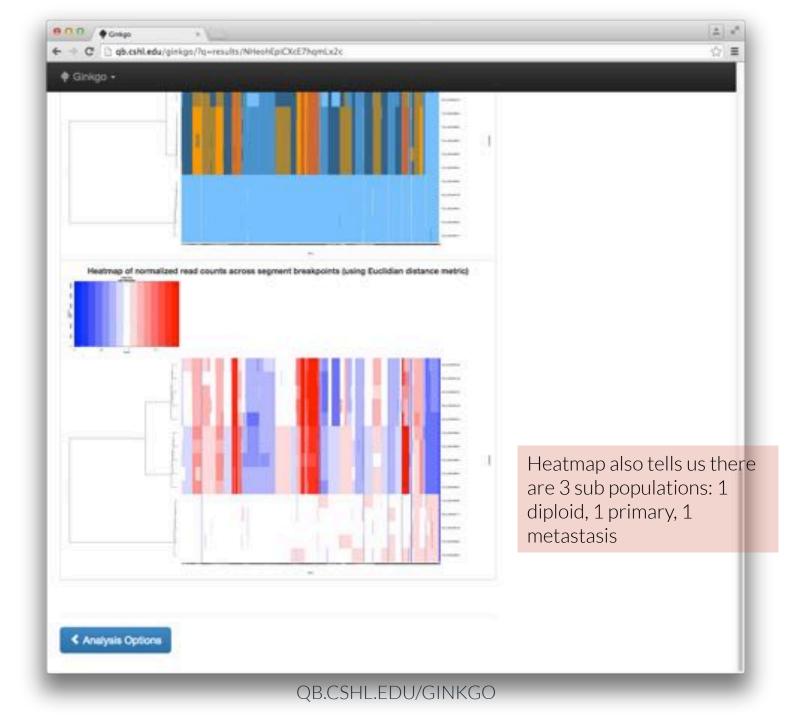
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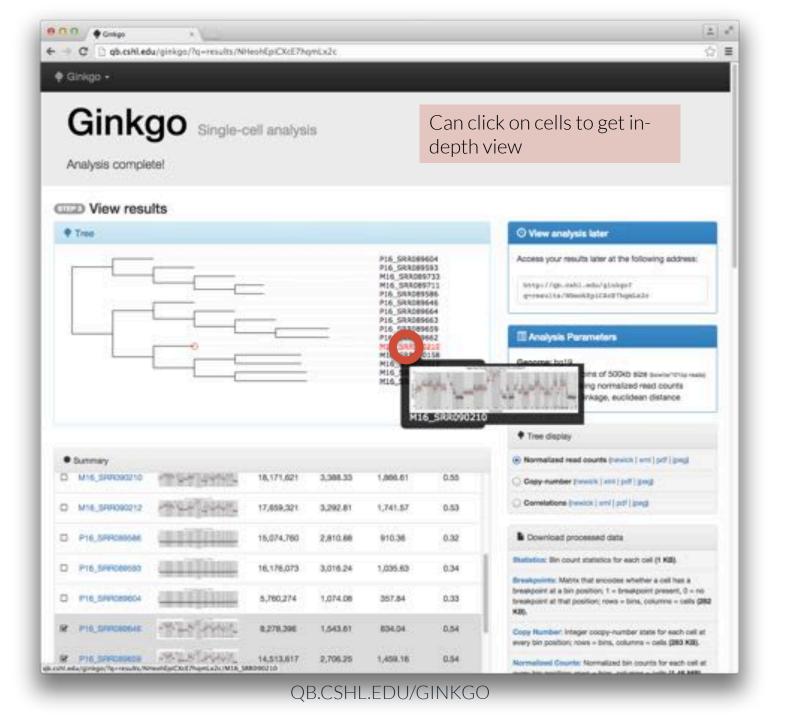
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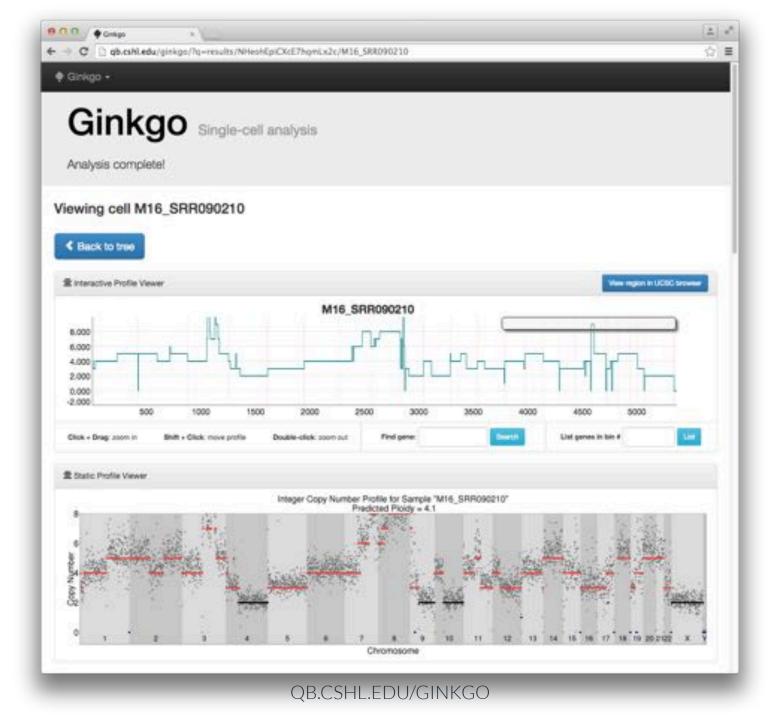
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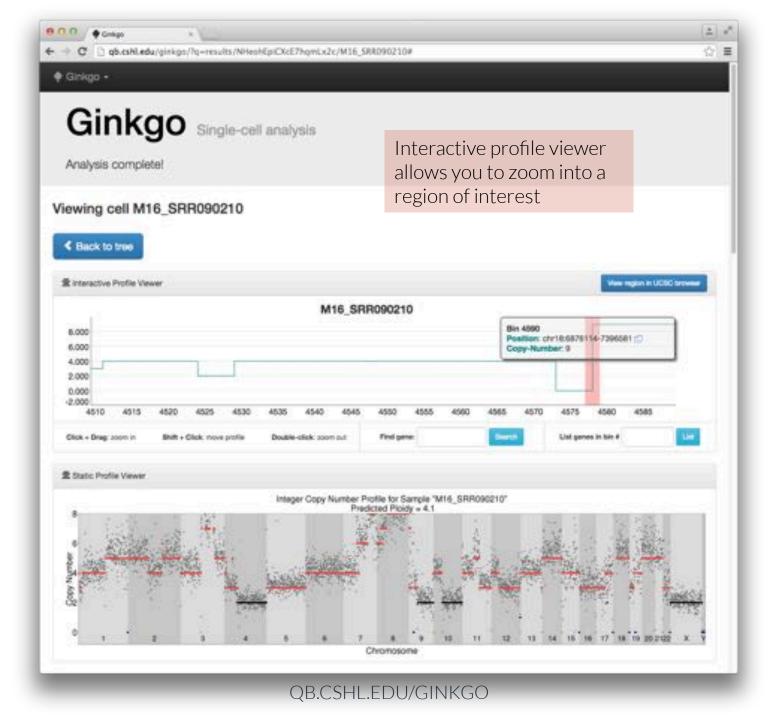
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M16_5HR090210     M16_5HR090212     P16_5HR090212	contraction of the second	17,659,321	3,292.61	1,741.57	0.53	Normalized read counts (newtok) emil (pdf) (peg)     Copy-number (newtok ( emil (edi ) peg)     Correlations (news) ( emil (edi ) peg)     Download processed data     Referice: Dir count statistics for each cel (1 KB).
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Mite_statesectro     Mite_statesectro     Mite_statesectro     Prie_statesecte     Prie_statesecte		17,859,321 15,074,760 16,176,073	3,292,61 2,810,68 3,016,24	1,741.57 910.36 1,035.63	0.53 0.32 0.34	Hormatipad read counts (newtok) emil (pdf) (peg)     Cegy-number (newtok) emil (pdf) (peg)     Correlations (newtok) emil (pdf) (peg)     Dovertoad processed data     Betenform: Sin count statistics for each cell (1 KB).     Breakpoint at a tim position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at that position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the position; 1 = breakpoint present, 0 = no breakpoint at the posint present present present present present present present presen
M16_5999090210     M16_599090212     P16_599099096     P16_599099096     P16_599099004		17,859,321 15,074,760 16,176,073 5,760,274	3.292.81 2.810.88 3.016.24 1.074.08	1,741.57 910.36 1,035.63 357.84	0.53 0.32 0.34 0.33	Hormalized read counts (newtok   smi   pdf   peg)     Capy-number (newtok   smi   pdf   peg)     Genelations (newtok   smi   pdf   peg)     Dovertoad processed data     Batistics: Bin count statistics for each cel (1 KB).     Breakpoint at a bin position; 1 - breakpoint present, 0 - no     breakpoint at a bin position; 1 - breakpoint present, 0 - no     breakpoint at a bin position; 1 - breakpoint present, 0 - no     breakpoint at a bin position; 1 - breakpoint present, 0 - no     breakpoint at a bin position; 1 - breakpoint present, 0 - no     breakpoint at a bin position; 1 - breakpoint present, 0 - no     breakpoint at a bin position; 1 - breakpoint present, 0 - no     breakpoint at their position; news - bins, columns - cells [282     KB).

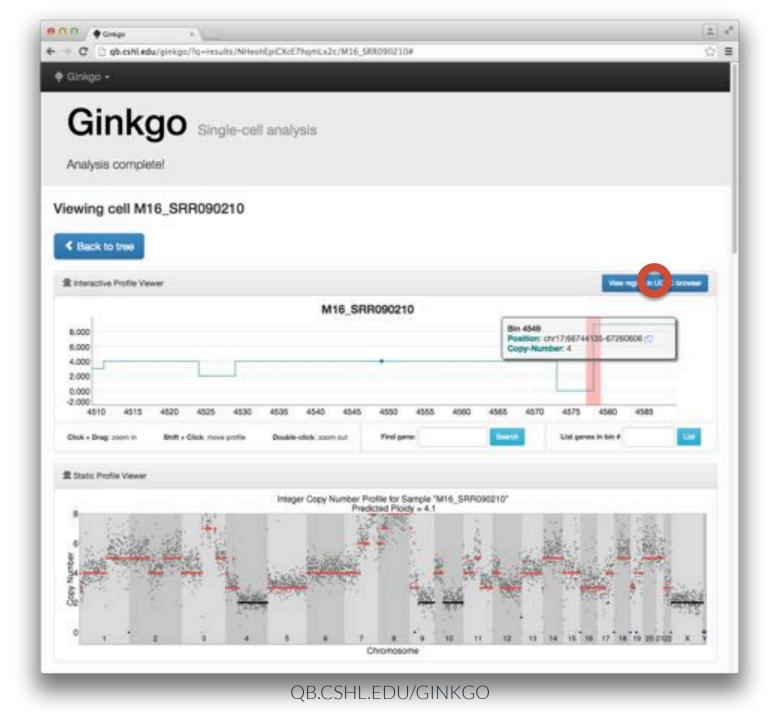


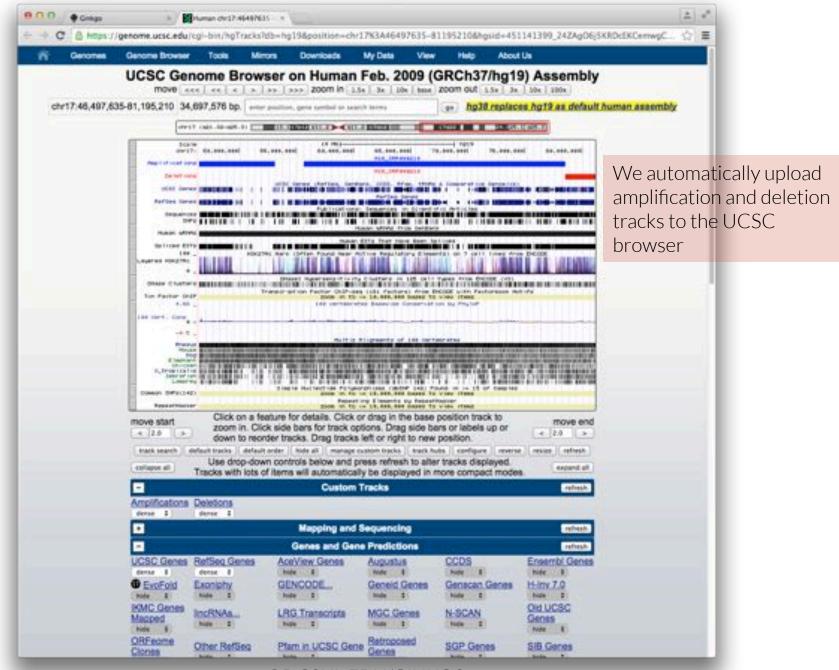




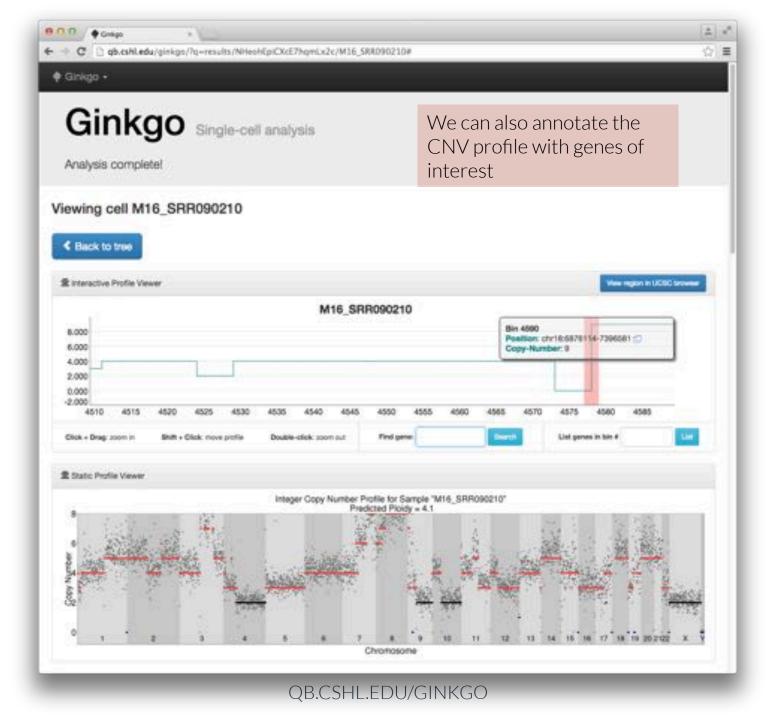


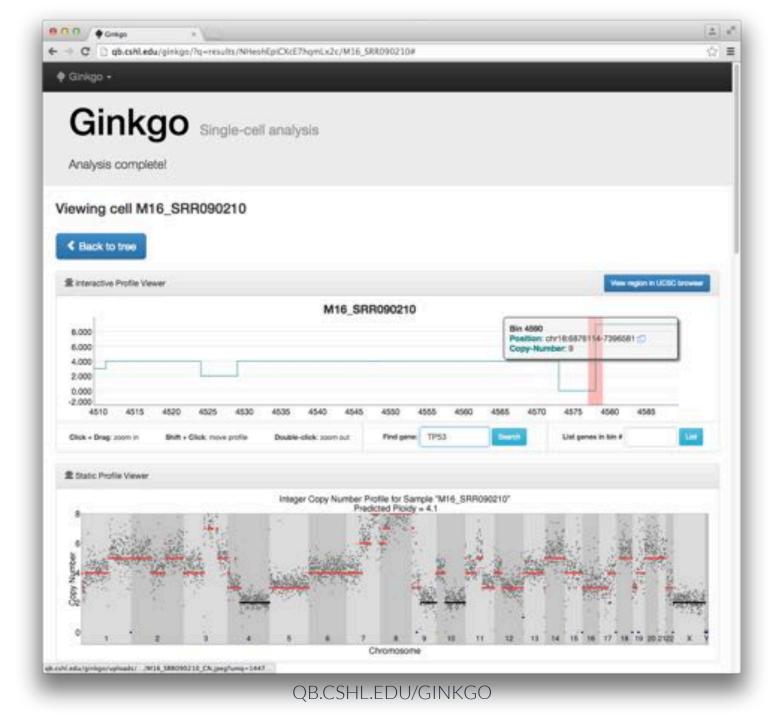






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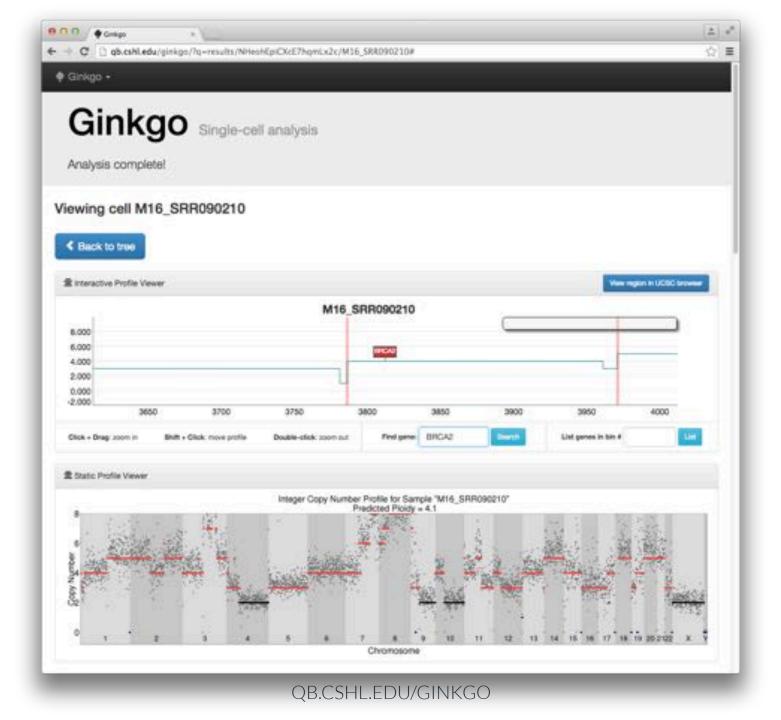




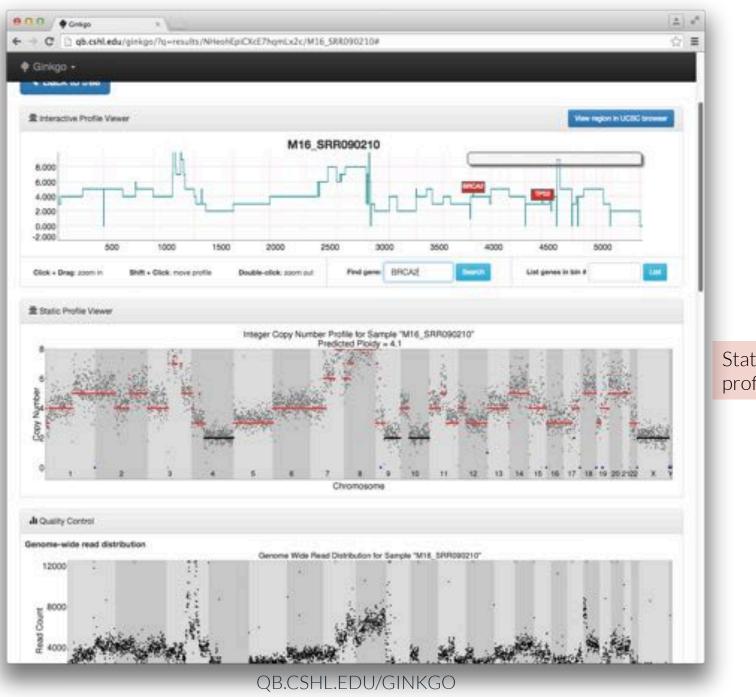




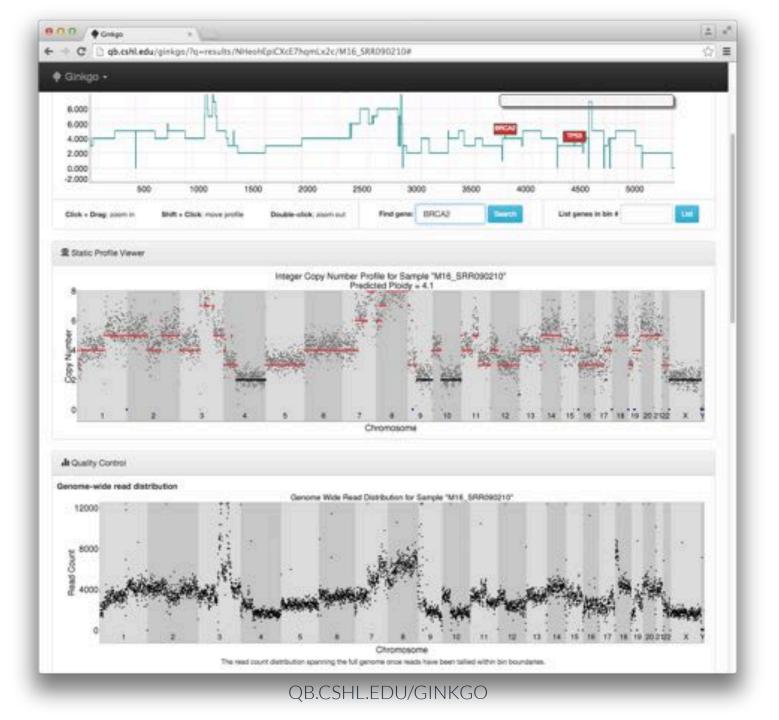


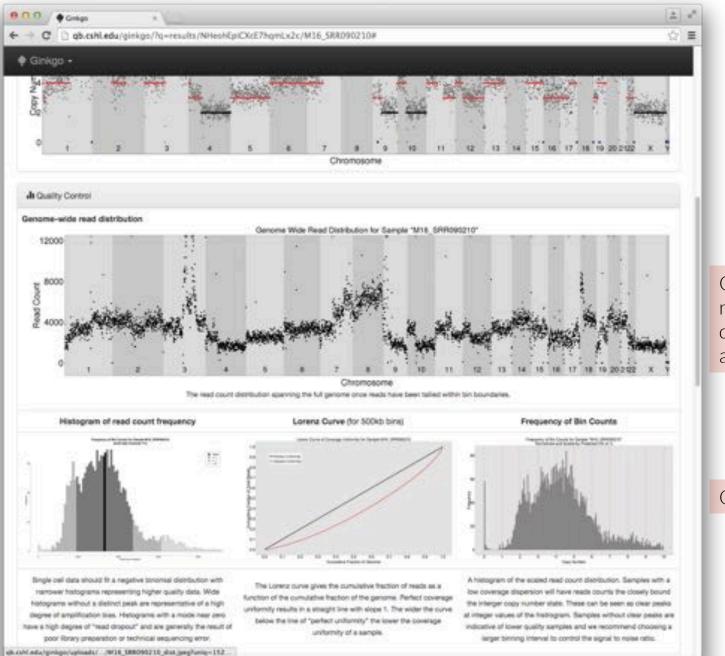






Static CNV profile

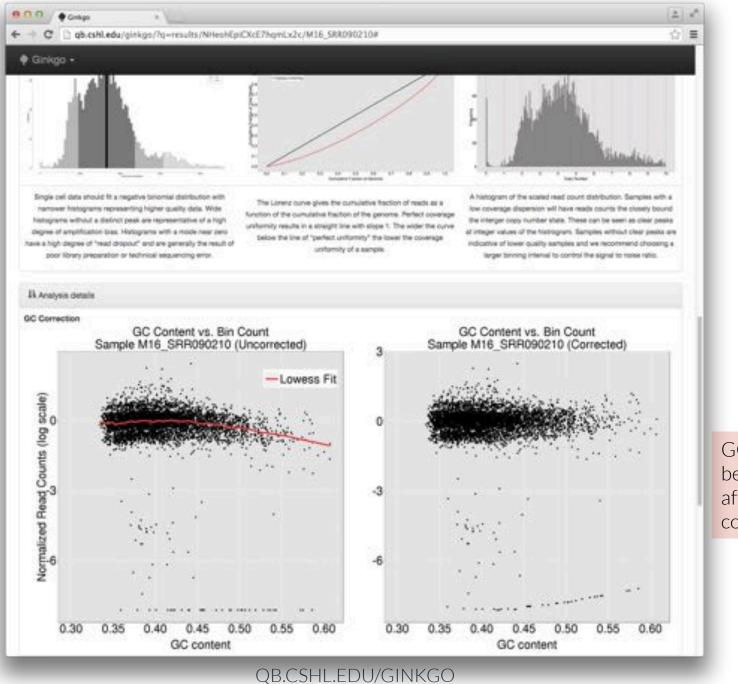




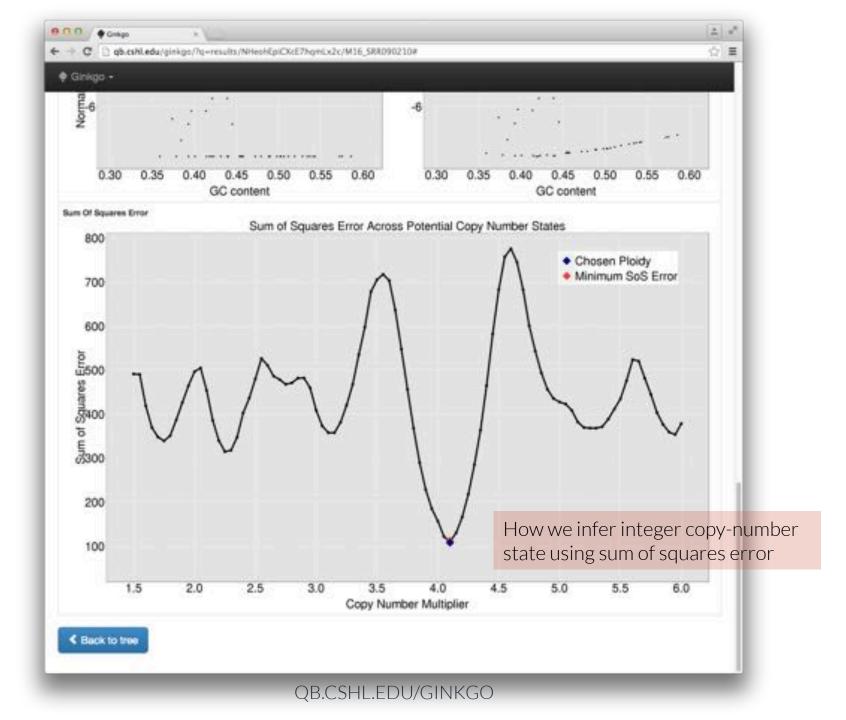
QB.CSHL.EDU/GINKGO

Original read count data before analysis

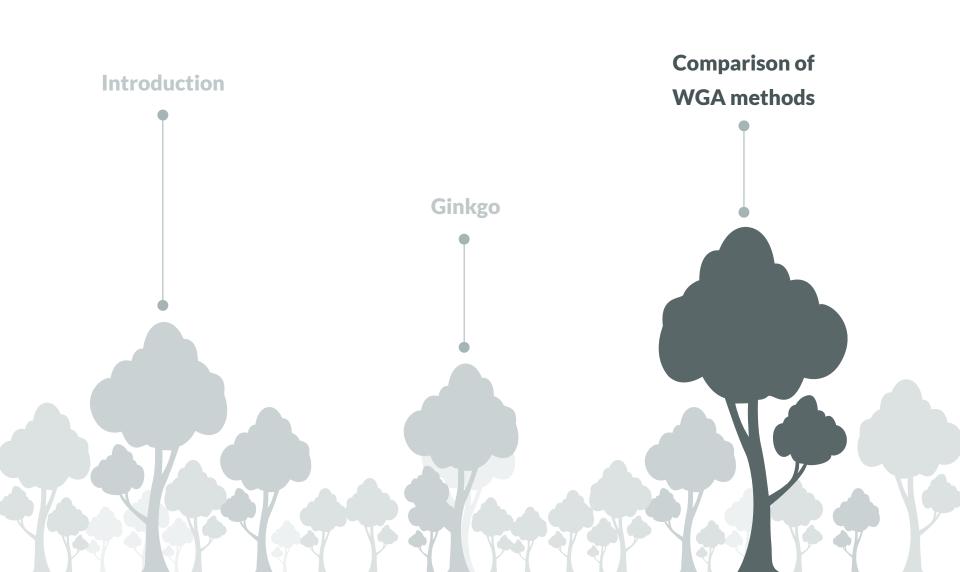
#### QC metrics



GC bias, before and after correction



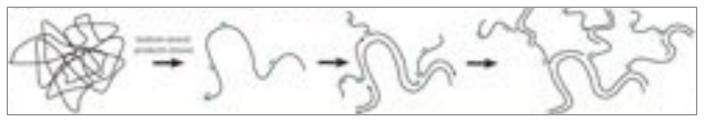
# Outline



# Whole Genome Amplification (WGA) methods



DOP-PCR (Degenerate Oligonucleotide Primed PCR)



MDA (Multiple Displacement Amplification)



MALBAC (Multiple Annealing and Looping Based Amplification Cycles)

# Comparison of WGA methods

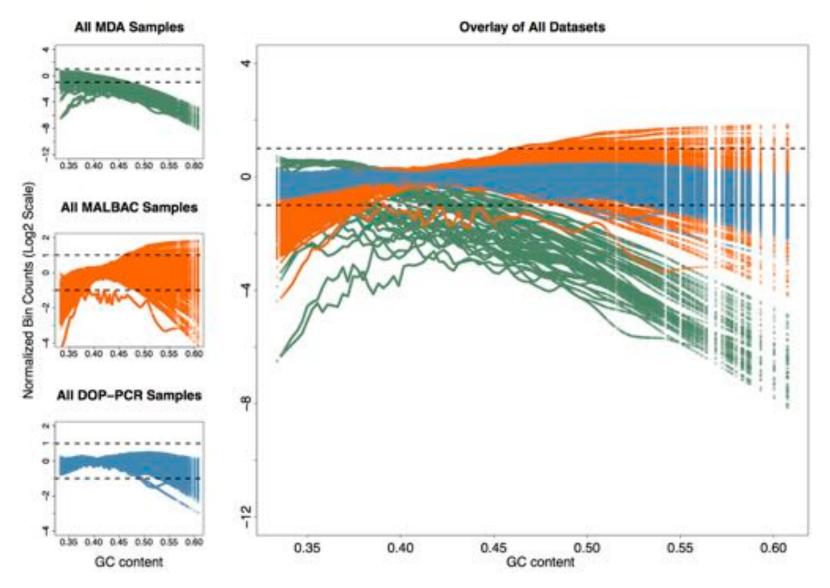
Paper	WGA Method	Tissue
Navin et al., 2011	DOP-PCR	Breast (T10)
Navin et al., 2011	DOP-PCR	Breast (T16P/M)
McConnnell et al., 2013	DOP-PCR	Neuron
Lu et al., 2012	MALBAC	Sperm
Ni et al., 2013	MALBAC	Lung
Hou et al., 2013	MALBAC	Oocyte
Kirkness et al., 2013	MDA	Sperm
Wang et al., 2012	MDA	Sperm
Evrony et al., 2012	MDA	Neuron

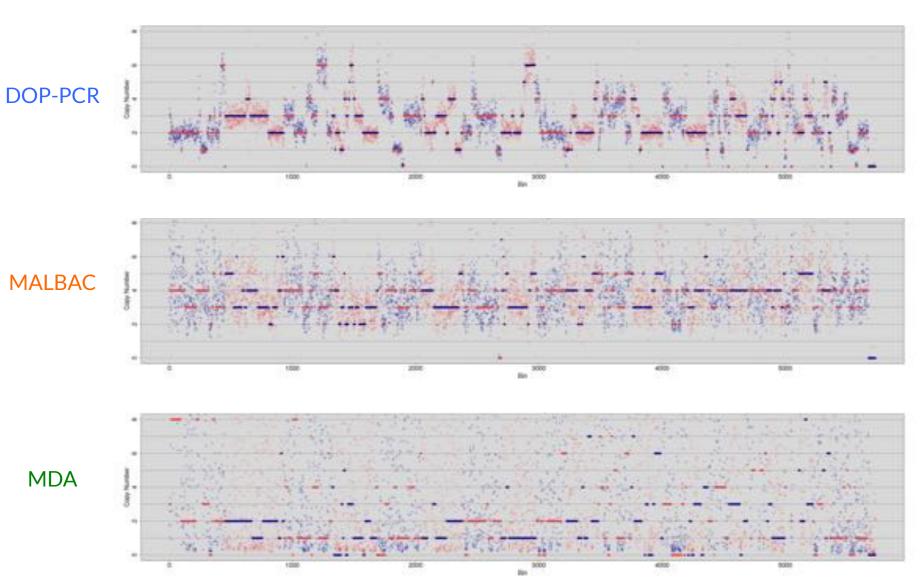
Explore the effects of WGA method on data quality:

- 1) GC bias
- 2) Coverage dispersion

Garvin and Aboukhalil et al., Nature Methods, 2015

GC Bias





Coverage Dispersion

Garvin and Aboukhalil et al., Nature Methods, 2015

### Coverage Dispersion

#### 1.0 **DOP-PCR** MALBAC **MDA** Median Absolute Deviation (MAD) 0.8 0.6 0.4 0.2 0.0 ž Hou З Wang Evrony McConnell Kirkness Navin (T10) Vavin (T16)

Median Absolute Deviation of Neighboring Bins

 $MAD = median_i \left( |X_i - median_j(X_j)| \right),$ 

# Summary

- Ginkgo is a platform for single-cell CNV analysis and visualization
- For copy-number analysis, we recommend DOP-PCR
- Check out Ginkgo and give us feedback
  - qb.cshl.edu/ginkgo
  - Garvin and Aboukhalil et al., Nature Methods, 2015



21,500 **PAGEVIEWS** 





Nov 3, San Francisco

### Thanks

#### **Ginkgo Team**

**Tyler Garvin** Jude Kendall Timour Baslan Jim Hicks Gurinder S. Atwal Michael Wigler Michael C. Schatz

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