Genome Arithmetic

Michael Schatz

Feb 17, 2020 Lecture 7: Applied Comparative Genomics



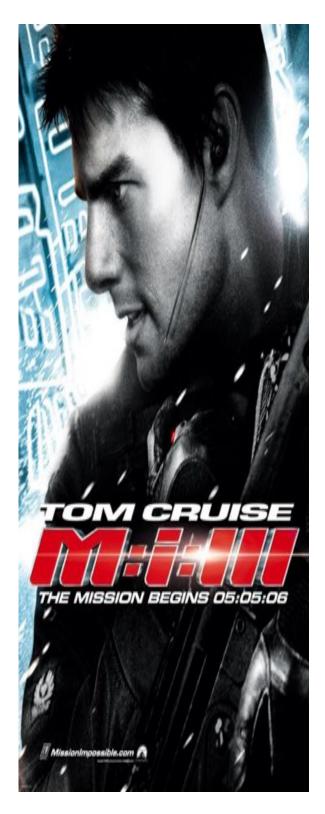
Assignment 2: Genome Assembly Due Wednesday Feb 12 @ 11:59pm

- I. Setup Docker/Ubuntu
- 2. Initialize Tools
- 3. Download Reference Genome & Reads

4. Decode the secret message

- I. Estimate coverage, check read quality
- 2. Check kmer distribution
- 3. Assemble the reads with spades
- 4. Align to reference with MUMmer
- 5. Extract foreign sequence
- 6. dna-encode.pl -d

https://github.com/schatzlab/appliedgenomics2020/blob/mas ter/assignments/assignment2/README.md



Assignment 3: Due Wed Feb 19

Assignment 3: Coverage, Genome Assembly, and Variant Calling

Assignment Date: Wednesday, Feb. 12, 2020 Due Date: Wednesday, Feb. 19, 2020 @ 11:59pm

Some of the tools you will need to use only run in a linux or mac environment. If you do not have access to a linux/mac machine, download and install a virtual machine or ubuntu instance following the directions here: https://github.com/schatzlab/sppliedgenomics2018/blob/master/assignments/virtualbox.md

Alternatively, you might also want to try out this docker instance that has these tools preinstalled: https://github.com/mschatz/wga-essentials

Question 1. Coverage simulator [10 pts]

- Q1a. How many 100bp reads are needed to sequence a 1Mbp genome to 5x coverage?
- Q1b. In the language of your choice, simulate sequencing 5x coverage of a 1Mbp genome and plot the histogram of coverage. Note you do not need to actually output the sequences of the reads, you can just randomly sample positions in the genome and record the coverage. You do not need to consider the strand of each read. The start position of each read should have a uniform random probability at each possible starting position (1 through 999,900). You can record the coverage in an array of 1M positions. Overlay the histogram with a Poisson distribution with lambda=5
- Q1c. Using the histogram from 1b, how much of the genome has not been sequenced (has 0x coverage). How well does this match Poisson expectations?
- Q1d. Now repeat the analysis with 15x coverage: 1. simulate the appropriate number of reads, 2. make a histogram, 3. overlay a Poisson distribution with tambda=15, 4. compute the number
 of bases with 0x coverage, and 5. evaluated how well it matches the Poisson expectation.

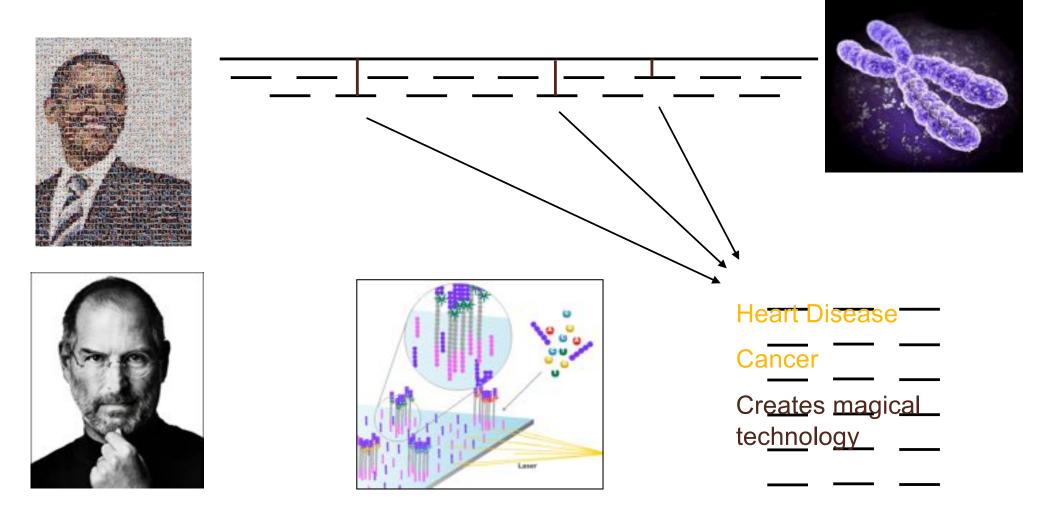
Question 2. de Bruijn Graph construction [10 pts]

Q2a. Draw (by hand or by code) the de Bruijn graph for the following reads using k+3 (assume all reads are from the forward strand, no sequencing errors, complete coverage of the genome)

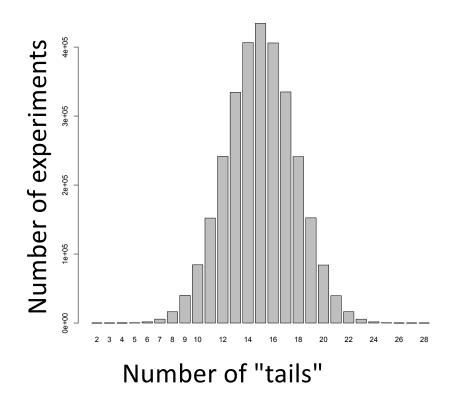
ATTCA ATTCA CATTC CATTC CATTC GATTC TATTT

Personal Genomics

How does your genome compare to the reference?



So, with 30 tosses (reads), we are much more likely to see an even mix of alternate and reference alleles at a heterozygous locus in a genome



This is why <u>at least</u> a "30X" (30 fold sequence coverage) genome is recommended: it confers sufficient power to distinguish heterozygous alleles and from mere sequencing errors

P(3/30 het) <?> P(3/30 err)

Thinking about allele sampling with the binomial distribution

The **binomial distribution** with parameters *n* and *p* is the <u>discrete probability</u> <u>distribution</u> of the number of successes in a sequence of <u>*n* independent yes</u> (e.g., "heads" or "reference allele") or <u>no</u> (e.g., "tails", or "alternate allele") experiments, <u>each of which yields success with probability p</u>.

The probability of getting exactly k successes in n trials is given by the probability mass function:

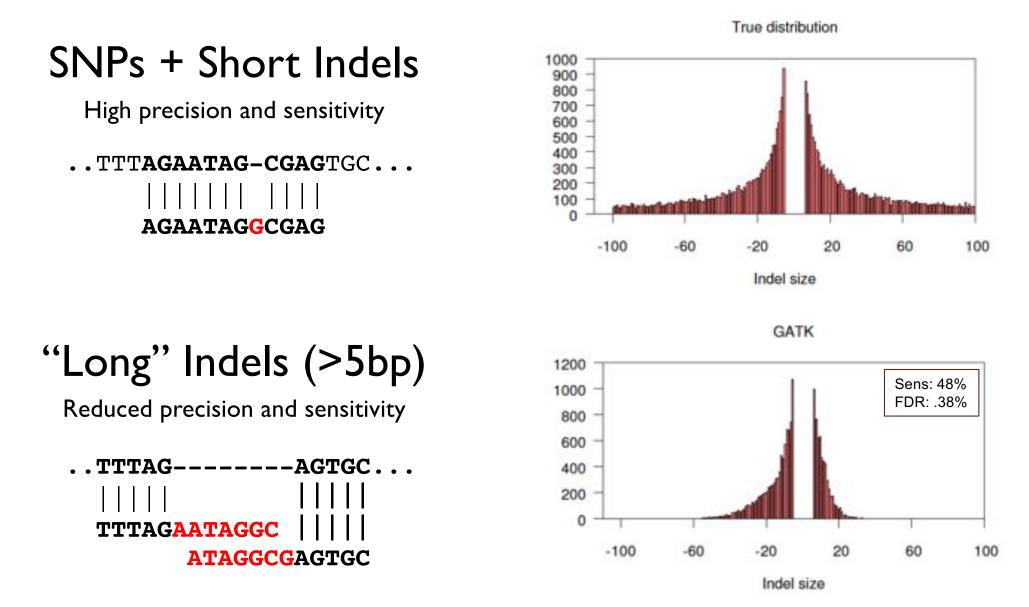
$$\Pr(X=k) = inom{n}{k} p^k (1-p)^{n-k}$$

What is the probability of seeing k=1 tails in n=3 flips of a fair coin with the probability of a tail (p) = 0.5?

3 choose 1 = 3; 0.5¹ = 0.5; (1-0.5)⁽³⁻¹⁾ = 0.25. So.... 3*0.5*0.25 = **0.375**

In R, the function would be: dbinom(1, size=3, prob=0.5)

Variation Detection Complexity



Analysis confounded by sequencing errors, localized repeats, allele biases, and mismapped reads

Scalpel: Haplotype Microassembly

DNA sequence **micro-assembly** pipeline for accurate detection and validation of *de novo* mutations (SNPs, indels) within exome-capture data.

Features

- I. Combine mapping and assembly
- 2. Exhaustive search of haplotypes
- 3. De novo mutations

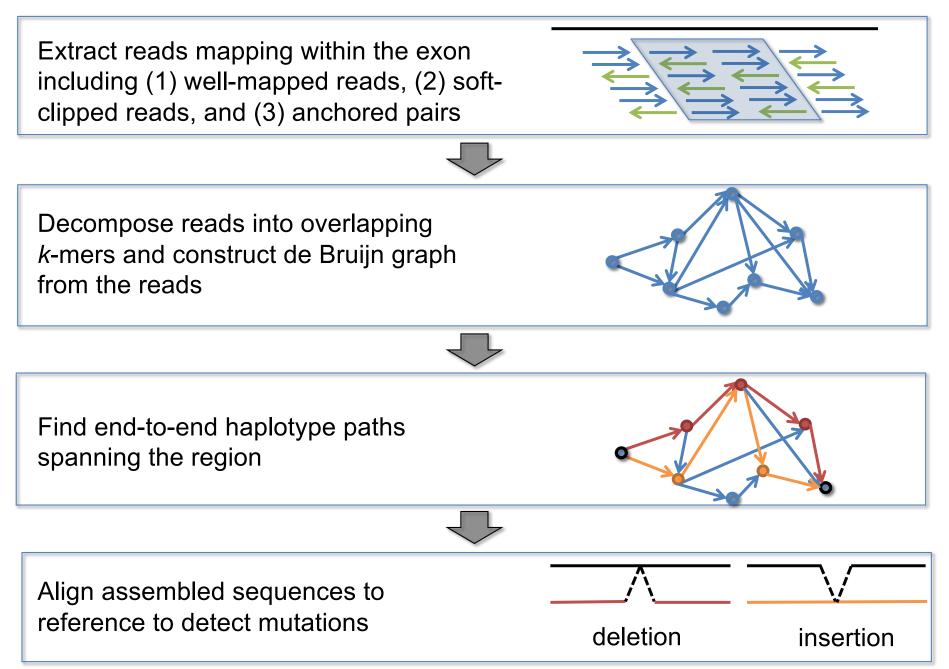


NRXN1 *de novo* SNP (auSSC12501 chr2:50724605)

Accurate de novo and transmitted indel detection in exome-capture data using microassembly. Narzisi et al. (2014) Nature Methods. doi:10.1038/nmeth.3069



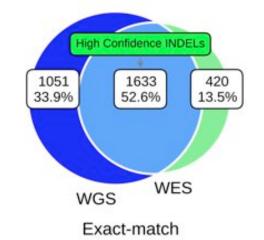
Scalpel Algorithm



Refined indel analysis

Examine sources of indel errors

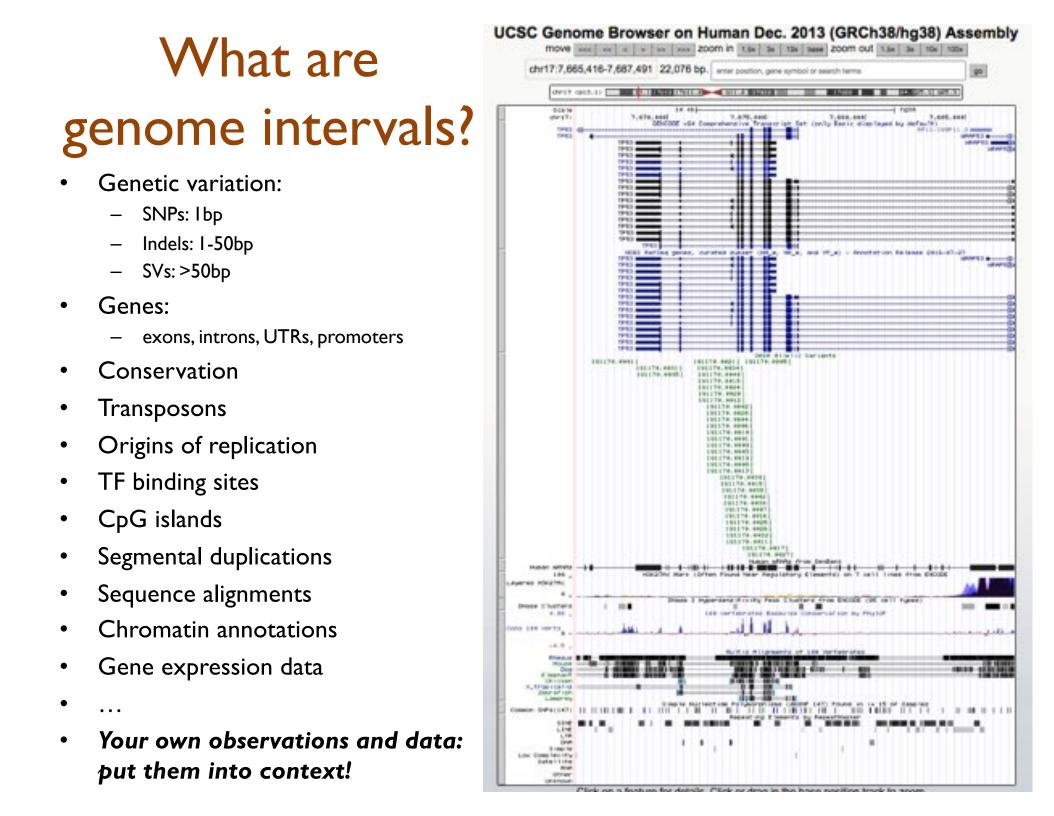
- Experimental validation of indels called from 30x whole genome vs. 110x whole exome of the same sample
- Most of the errors due to short microsatellite errors introduced during exome capture, also misses most long indels

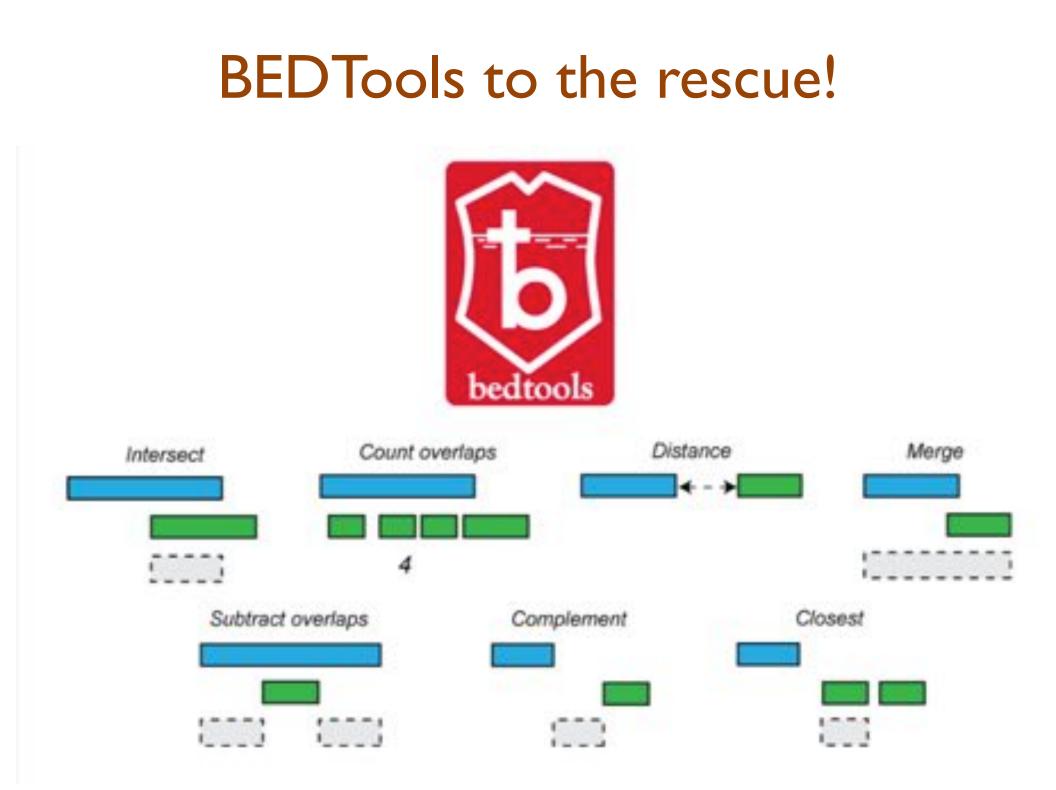


• Recommend WGS for indel analysis instead

	All INDELs	Valid	PPV	INDELs >5bp	Valid (>5bp)	PPV (>5bp)
Intersection	160	152	95.0%	18	18	100%
WGS	145	122	84.1%	33	25	75.8%
WES	161	91	56.5%	I	I	100%

Reducing INDEL calling errors in whole-genome and exome sequencing data Fang et al. Genome Medicine (2014) 6:89. doi:10.1186/s13073-014-0089-z





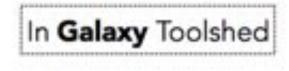
Getting & Using BEDTools

Run Batch Script	Help
Run iptools	
	Intersect Remove/Subtract Closest Window Coverage Multi-intersect Set path to REDTools

Integrated into IGV

BEDTools

- Intersect BAM alignments with intervals in another files
- Count intervals in one file overlapping intervals in another file
- Create a histopram of genome coverage
- Create a BedGraph of genome. **EDVERAGE**
- Convert from BAM to BED.
- Merge BedGraph files
- Intersect multiple sorted BED files



bedtools: a powerful toolser (x) a o o o a A o - - - 0 0 0 0 0 1 🎦 🛅 📴 JAUMAE Elli Dady 🔛 😰 💓 🏠 Elli schatzate 🔿 🔁 🖧 51. Elli cent Elli Av Elli Media 🔅 edit 🔅 Remove NYT Cookee III E: Other Bookmarks bectools v2.26.0 > next.



Bedtools is a fast, flexible toolset for genome arithmetic.

Bedtools links

Issue Tracker Source @ GitHub Old Releases @ Google Code Mailing list @ Google Groups Queries @ Biostar Quinlan lab @ UU

Sources

Browse source @ GitHub .

Releases

Stable releases now @ Github

This Page

Show Source

Quick search

Go

Enter search terms or a module.

Table of contents

bedtools: a powerful toolset for genome arithmetic

Collectively, the bedtools utilities are a swiss-army knife of tools for a wide-range of genomics analysis tasks. The most widely-used tools enable genome anthmetic: that is, set theory on the genome. For example, bedtools allows one to intersect, merge, count, complement, and shuffle genomic intervals from multiple files in widely-used genomic file formats such as BAM, BED, GFF/GTF, VCF. While each individual tool is designed to do a relatively simple task (e.g., intersect two interval files), guite sophisticated analyses can be conducted by combining multiple bedtools operations on the UNIX command line.

bedtools is developed in the Quinlan laboratory at the University of Utah and benefits from fantastic contributions made by scientists worldwide.

Tutorial

We have developed a fairly comprehensive tutorial that demonstrates both the basics, as well as some more advanced examples of how bedtools can help you in your research. Please have a look.

Interesting Usage Examples

In addition, here are a few examples of how bedtools has been used for genome research. If you have interesting examples, please send them our way and we will add them to the list.

- Coverage analysis for targeted DNA capture. Thanks to Stephen Turner.
- Measuring similarity of DNase hypersensitivity among many cell types
- Extracting promoter sequences from a genome
- Comparing intersections among many genome interval files
- · RNA-seq coverage analysis. Thanks to Erik Minikel.
- · Identifying targeted regions that lack coverage. Thanks to Brent Pedersen.
- Calculating GC content for CCDS exons.
- Making a master table of ChromHMM tracks for multiple cell types.

Extensive Documentation and Examples

BED Format

BED (Browser Extensible Data) format provides a flexible way to define intervals.

The first three required BED fields are:

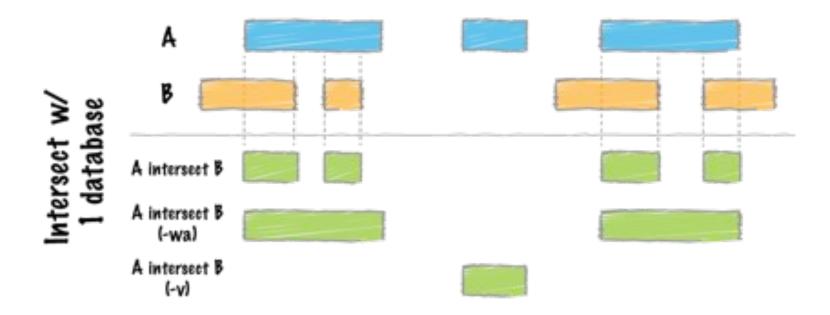
- 1. chrom The name of the chromosome (e.g. chr3, chrY, chr2_random) or scaffold (e.g. scaffold10671).
- 2. chromStart The starting position of the feature in the chromosome or scaffold. The first base in a sequence is numbered 0.
- 3. chromEnd The ending position of the feature in the chromosome or scaffold. The chromEnd base is not included in the display of the feature. For example, the first 100 bases of a chromosome are defined as chromStart=0, chromEnd=100, and span the bases numbered 0-99.

The 9 additional optional BED fields are:

- 1. name Defines the name of the BED line
- 2. score A score between 0 and 1000
- 3. strand Defines the strand. Either "." (=no strand) or "+" or "-".
- 4. thickStart The starting position at which the feature is drawn thickly
- 5. thickEnd The ending position at which the feature is drawn thickly (for example the stop codon in gene displays).
- 6. itemRgb An RGB value of the form R,G,B (e.g. 255,0,0).
- 7. blockCount The number of blocks (exons) in the BED line.
- 8. blockSizes A comma-separated list of the block sizes. The number of items in this list should correspond to blockCount.
- 9. blockStarts A comma-separated list of block starts. All of the blockStart positions should be calculated relative to chromStart. The number of items in this list should correspond to blockCount.

## gene	s.bed has: ch	rom, txStart,	txEnd,	name,	num_exons,	and strand
<pre>\$ head -n4 genes.bed</pre>						
chr1	134212701	134230065	Nuak2	8	+	
chr1	134212701	134230065	Nuak2	7	+	
chr1	33510655	33726603	Prim2,	14	_	
chr1	25124320	25886552	Bai3,	31	-	

BEDTools Intersect



What exons are hit by SVs?

\$ cat A.bed chr1 10 20 chr1 30 40 \$ cat B.bed chr1 15 20 \$ bedtools intersect -a A.bed -b B.bed -wa chr1 10 20

What parts of exons are hit by SVs?

\$ cat	A.be	d					
chr1	10 3	20					
chr1	30	40					
\$ cat	B.be	d					
chr1	15	20					
\$ bedt	tools	intersect	- a	A.bed	-b	B.bed	
chr1	15	20					

BEDTools Merge



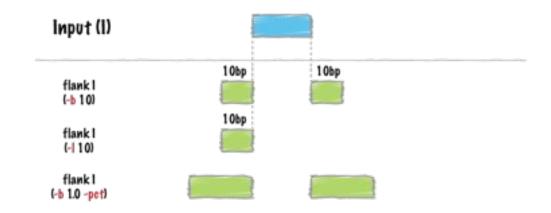
What parts of the genome are exonic?

bedtoo	ls merge	-i exons.bed	1	head	-n	20
chr1	11873	12227				
chr1	12612	12721				
chr1	13220	14829				
chr1	14969	15038				
chr1	15795	15947				
chr1	16606	16765				
chr1	16857	17055				
	47222	1770				

Note input must be sorted!

sort	-k1,1	-k2,2n	foo.bed	>	foo.sort.bed
------	-------	--------	---------	---	--------------

BEDTools Flank & getfasta



## gene	s.bed has: ch	rom, txStart,	txEnd,	name,	num_exons,	and strand	
\$ head	-n4 genes.bed						
chr1	134212701	134230065	Nuak2	8	+		
chr1	134212701	134230065	Nuak2	7	+		
chr1	33510655	33726603	Prim2,	14	-		
chr1	25124320	25886552	Bai3,	31	-		

Identify promoter regions (2kbp upstream)
\$ bedtools flank -i genes.bed -g mm9.chromsizes -1 2000 -r 0 -s > genes.2kb.promoters.bed

8

7

+

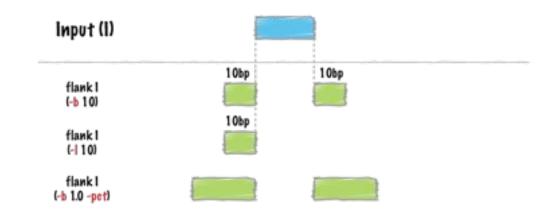
Т

Show promoter coordinates
\$ head genes.2kb.promoters.bed
chr1 134210701 134212701 Nuak2
chr1 134210701 134212701 Nuak2

	134210701	134212/01	NUARZ	/	•
chr1	33726603	33728603	Prim2,	14	_
chr1	25886552	25888552	Bai3,	31	_

Extract the sequences
\$ bedtools getfasta -fi mm9.fa -bed genes.2kb.promoters.bed -fo genes.2kb.promoters.bed.fa

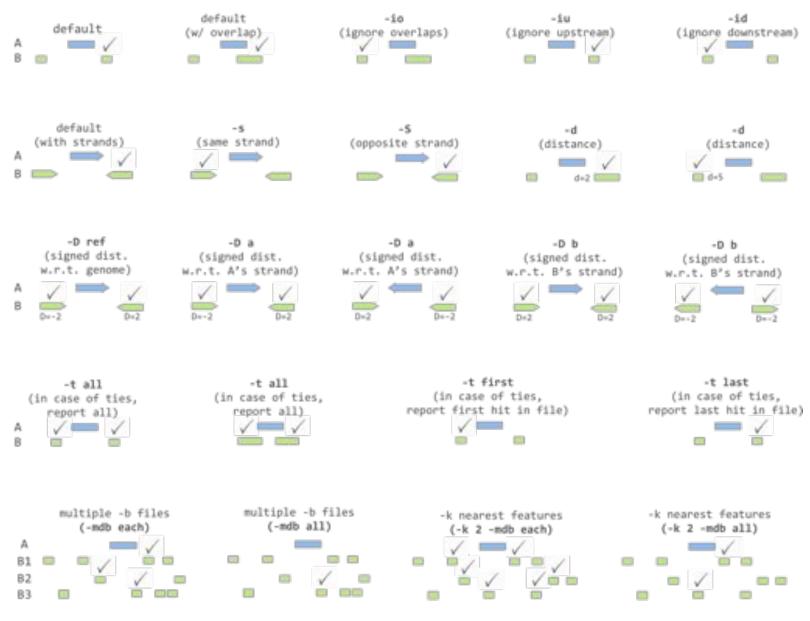
BEDTools Flank & getfasta



## gen	es.bed has: c	hrom, txStart	, txEnd,	name,	num_	exons,	and	strand
\$ head	l -n4 genes.be	ed						
chr1	134212701	134230065	Nuak2	8	+			
chr1	134212701	134230065	Nuak2	7	+			
chr1	33510655	33726603	Prim2,	14	-			
chr1	25124320	25886552	Bai3,	31	-			
<pre>## Identify promoter regions (2kbp upstream) \$ bedtools flank -i genes.bed -g mm9.chromsizes -1 2000 -r 0 -s > genes.2kb.promoters.bed</pre>								
## Show promoter coordinates Can also use the samtools faidx output								
\$ head	l genes.2kb.pr	omoters.bed	_					
chr1	134210701	134212701	Nuak2	8	+			
chr1	134210701	134212701	Nuak2	7	+			
chr1	33726603	33728603	Prim2,	14	-			
chr1	25886552	25888552	Bai3,	31	_			

Extract the sequences
\$ bedtools getfasta -fi mm9.fa -bed genes.2kb.promoters.bed -fo genes.2kb.promoters.bed.fa

BEDTools Closest



What is the gene closest to this SNP or this enhancer?

BEDTools commands

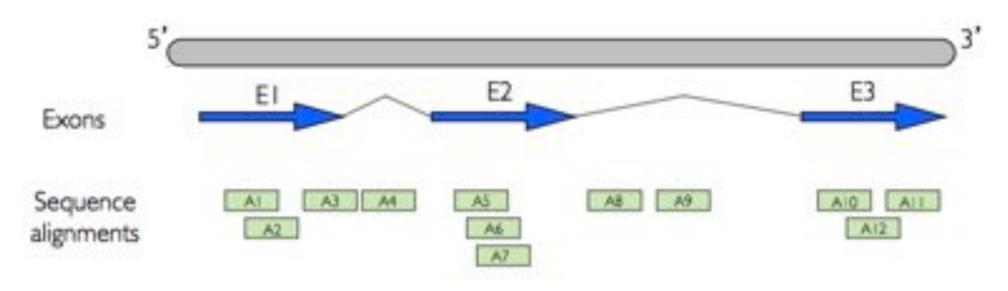
annotate bamtobed bamtofastq bed12tobed6 bedpetobam bedtobam closest cluster complement coverage expand flank fisher genomecov

getfasta groupby groupby igv intersect jaccard links makewindows map maskfasta merge multicov multiinter nuc

overlap pairtobed pairtopair random reldist shift shuffle slop sort subtract tag unionbedg window

http://bedtools.readthedocs.io/en/latest/content/bedtools-suite.html

BEDTools Performance



How many reads are aligned to exonic sequences?

```
$ awk '{if ($3=="exon"){print}}' gencode.v21.annotation.gff3 | wc -1
1162114
```

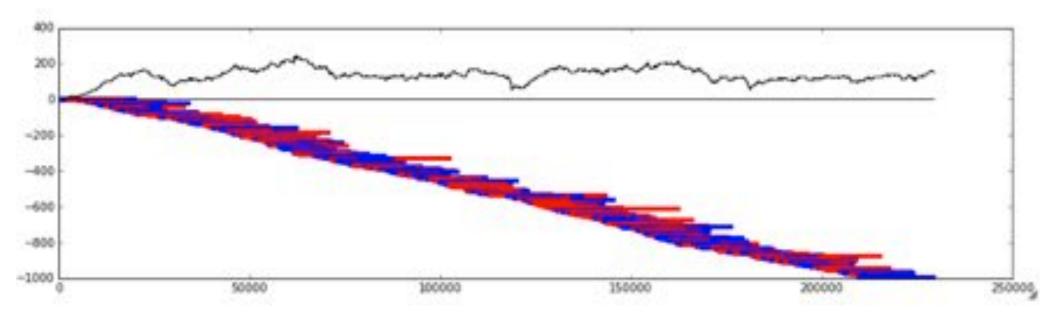
if ((read.start <= exon.end) && (read.end >= exon.start)) { print "in exon!"; }

How many comparison would a brute force approach take to scan a 30x dataset?

30x3Gb = 90Gbp / 100bp reads = 900M reads

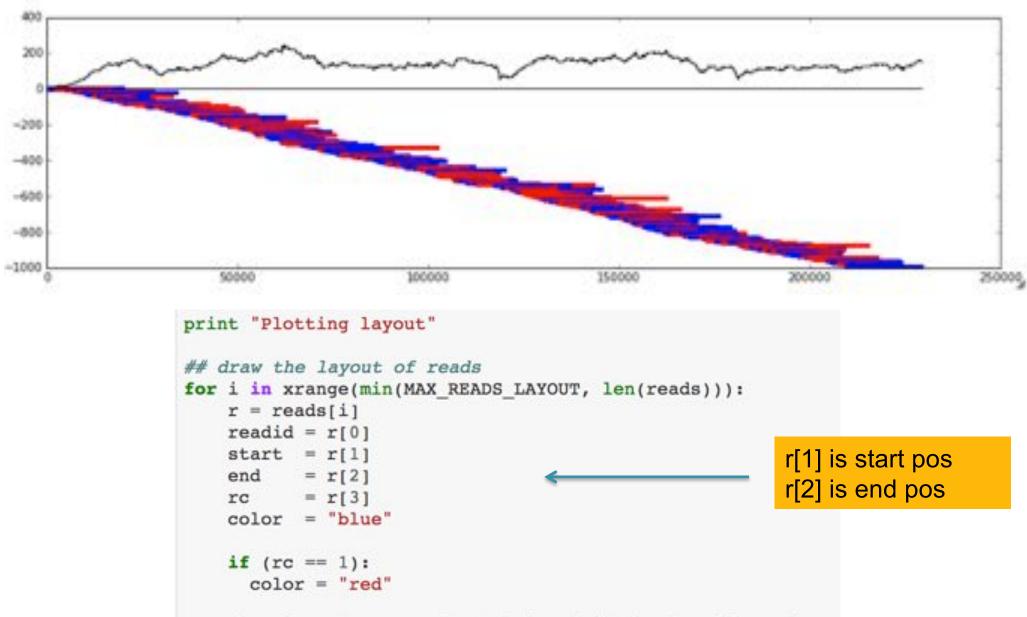
900M reads x 1.1M exons = 990MM comparisons! 😕

Coverage across the genome



\$ head -3 ~/readid.start.stop.txt
1 0 19814
2 799 19947
3 1844 13454
\$ tail -3 ~/readid.start.stop.txt
1871 973590 965902
1872 966703 973521
1873 973632 966946

Coverage across the genome



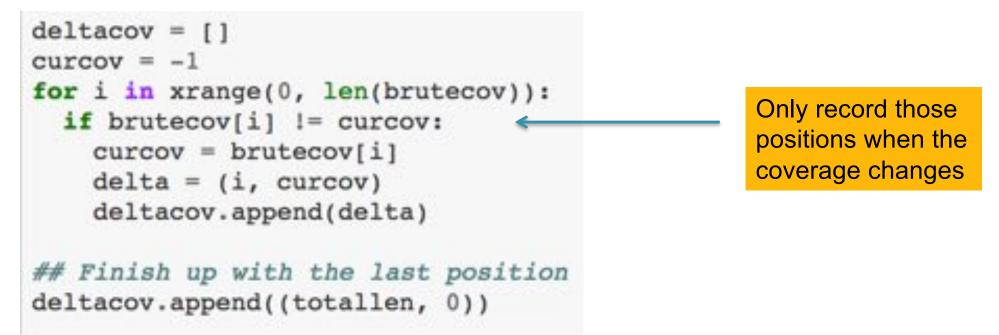
plt.plot ([start,end], [-2*i, -2*i], lw=4, color=color)

Brute Force Coverage Profile

```
print "Brute force computing coverage over %d bp" % (totallen)
starttime = time.time()
brutecov = [0] * totallen
for r in reads:
 # print " -- [%d, %d]" % (r[1], r[2])
                                                                Add 1 to coverage
                                                                vector at every
  for i in xrange(r[1], r[2]):
                                                                position the read
   brutecov[i] += 1
                                                                covers
brutetime = (time.time() - starttime) * 1000.0
print " Brute force complete in %0.02f ms" % (brutetime)
print brutecov[0:10]
                                                                * This is what you
                                                                should do for the
Brute force computing coverage over 973898 bp
  Brute force complete in 4435.00 ms
                                                                homework<sup>1</sup>*
[1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1]
```

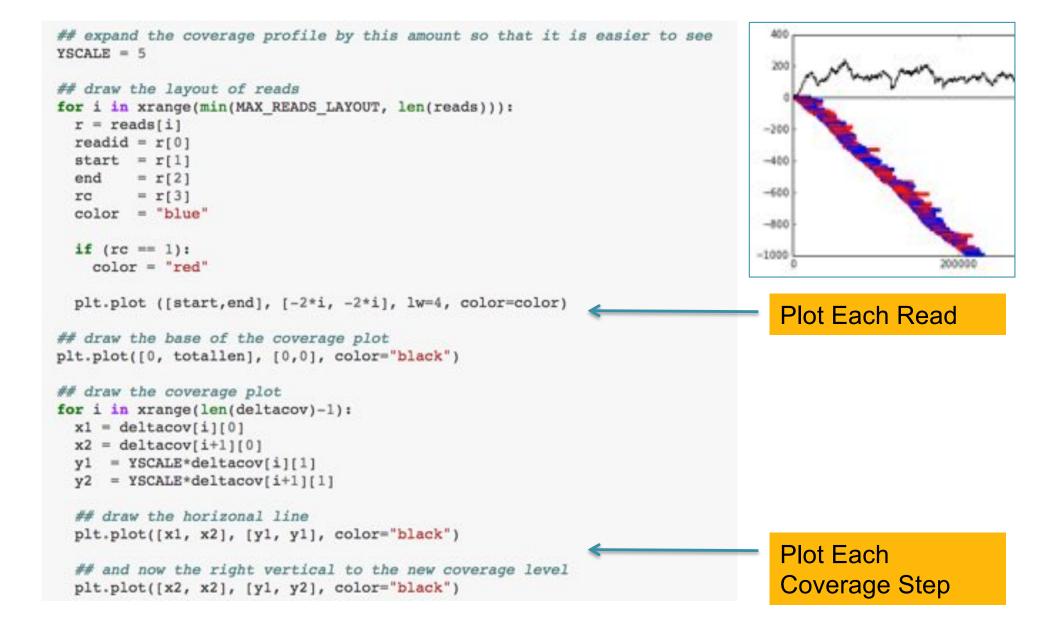
Notice that it took 4435 ms for this to complete

Delta Encoding aka run length encoding

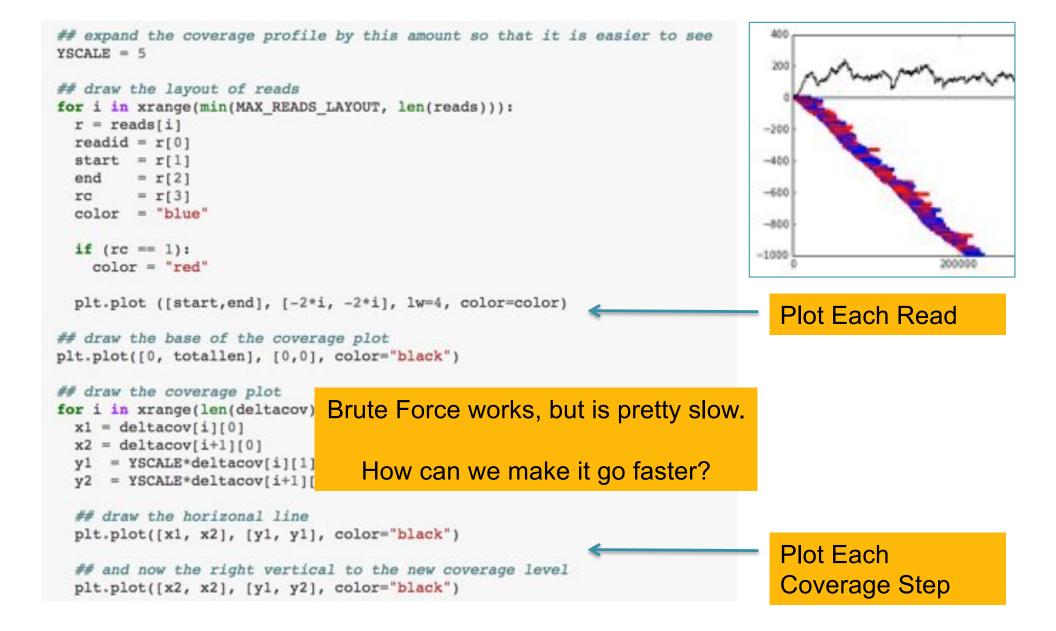


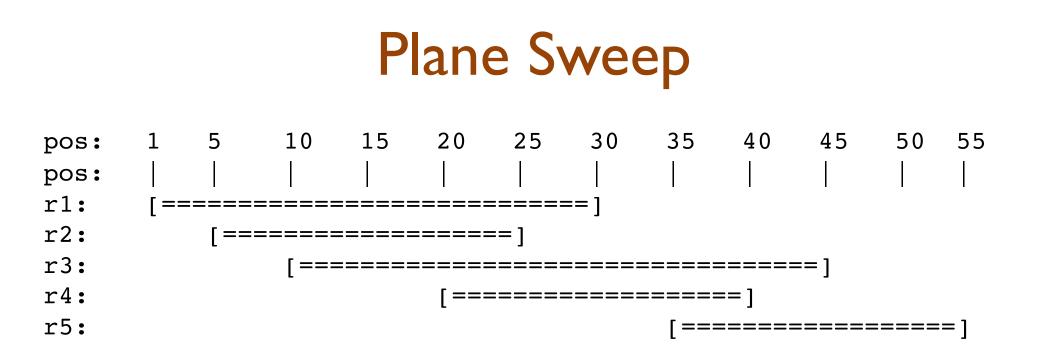
```
Delta encoding coverage plot
Delta encoding required only 3697 steps, saving 99.62% of the space in 151.32 ms
0: [0,1]
1: [799,2]
2: [1844,3]
...
3694: [973770,2]
3695: [973779,1]
3696: [973898,0]
```

Plot Coverage and Read Positions



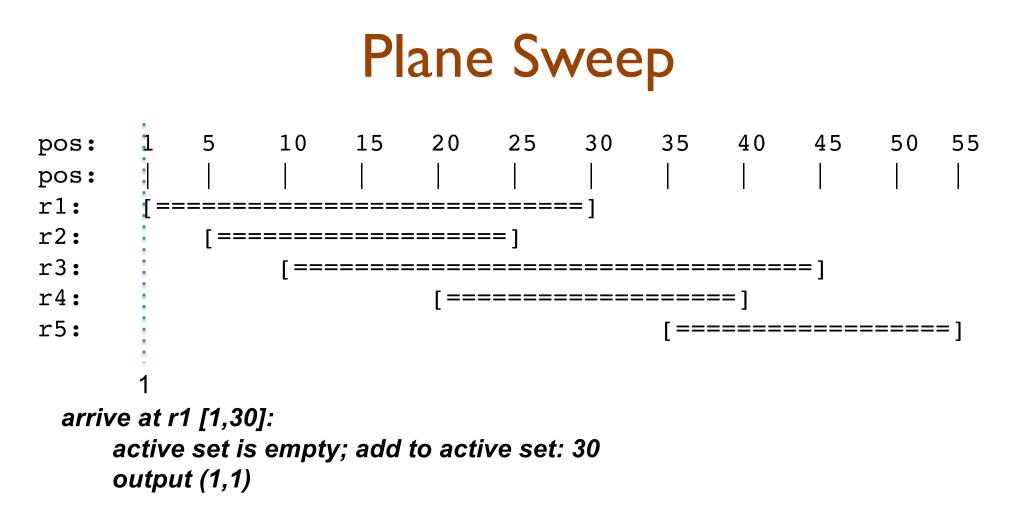
Plot Coverage and Read Positions

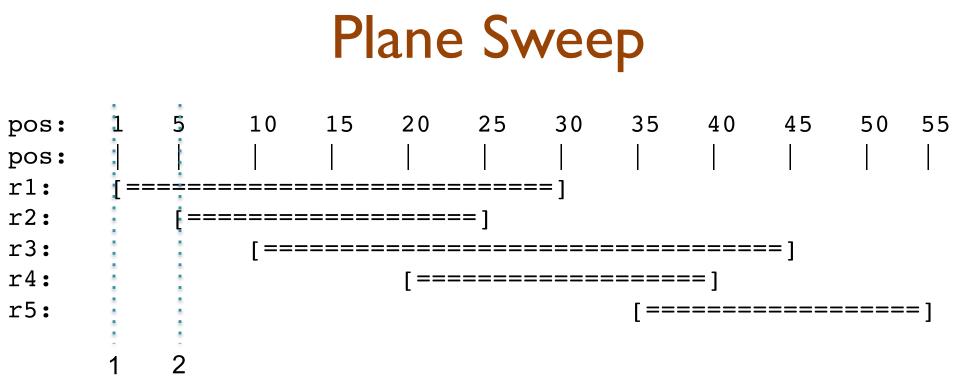




The basic algorithm works like this:

- Assume layout is in sorted order by start position (or explicitly sort by start position)
- use a "list" to track how many reads currently intersect the plane keyed by end coord
 - the number of elements in the list corresponds to the current depth
- walking from start position to start position
 - check to see if we past any read ends
 - coverage goes down by one when a read ends
 - coverage goes up by one when new read is encountered

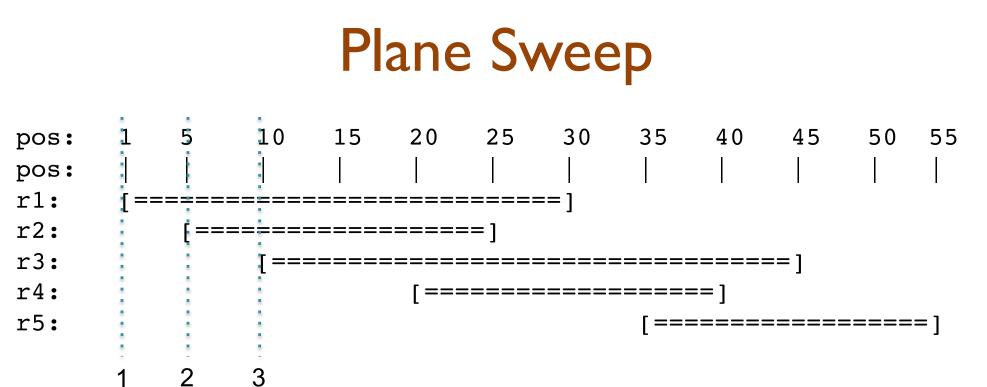




```
arrive at r1 [1,30]:
active set is empty; add to active set: 30
output (1,1)
```

```
arrive at r2 [5,25]:
```

5 < 30: add to active set: 25, 30 <- notice insert at beginning of active set output (5, 2)

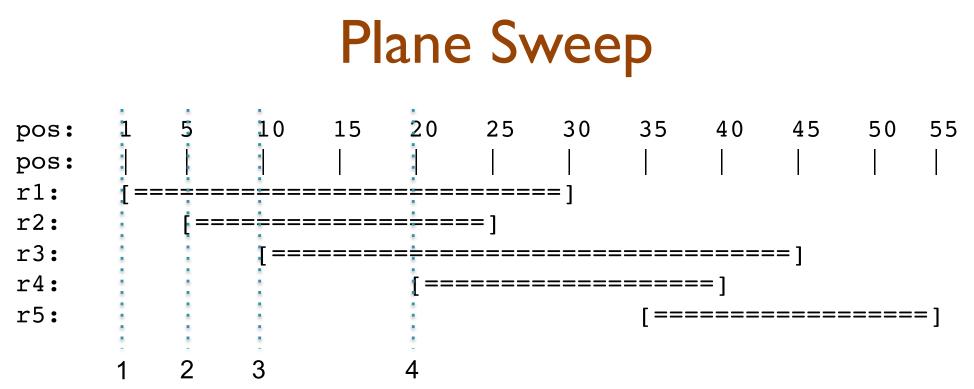


arrive at r1 [1,30]: active set is empty; add to active set: 30 output (1,1)

arrive at r2 [5,25]:

5 < 30: add 25 to active set: 25, 30 <- notice insert at beginning of active set output (5, 2)

```
arrive at r3 [10,45]:
10 < 25; add 45 to active set: 25, 30, 45 <- add to end of active set
output (10, 3)
```

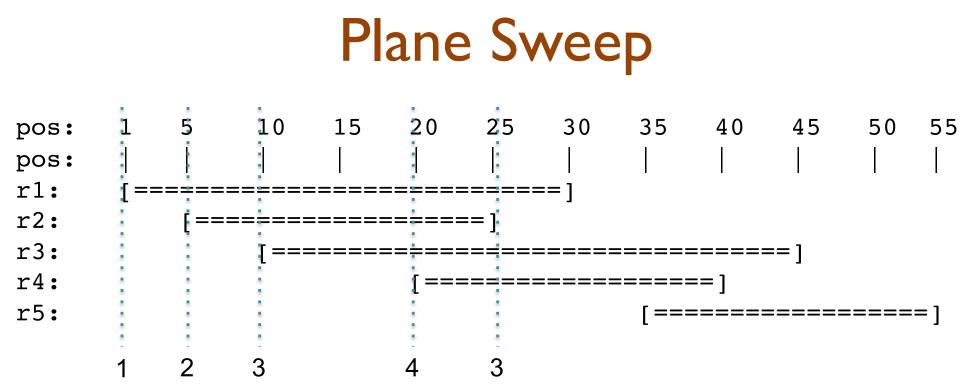


arrive at r3 [10,45]:

10 < 25; add 45 to active set: 25, 30, 45 <- add to end of active set output (10, 3)

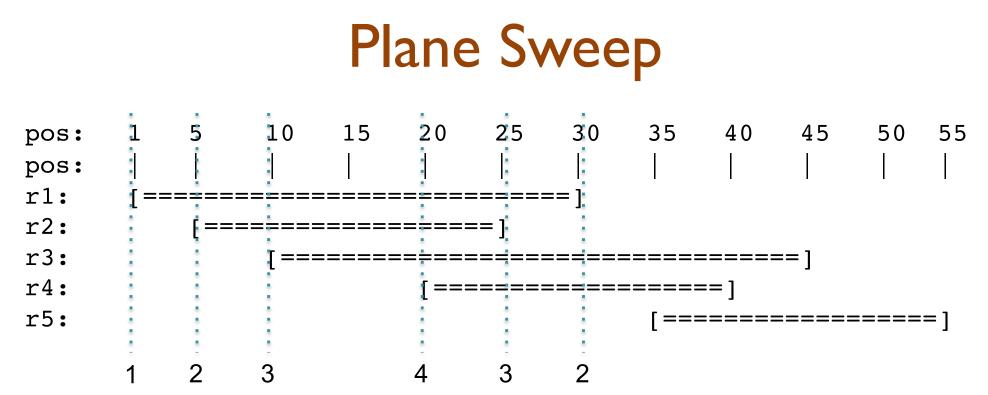
arrive at r4 [20,40]:

20 < 25; add 40 to active set: 25, 30, 40, 45 <- out of order again output (20, 4)



arrive at r5[35,55]:

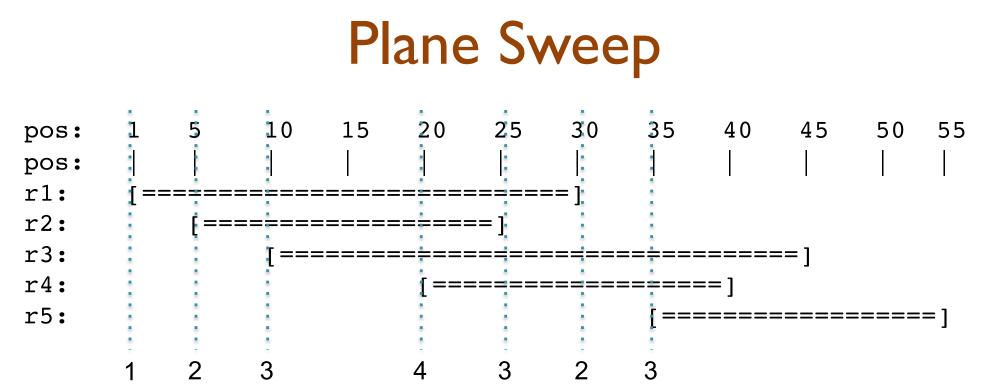
35 > 25: step down at 25; active set: 30, 40, 45 output (25, 3)



arrive at r5[35,55]:

35 > 25: step down at 25; active set: 30, 40, 45 output (25, 3)

35 > 30: step down at 30; active set: 40, 45 output (30, 2)

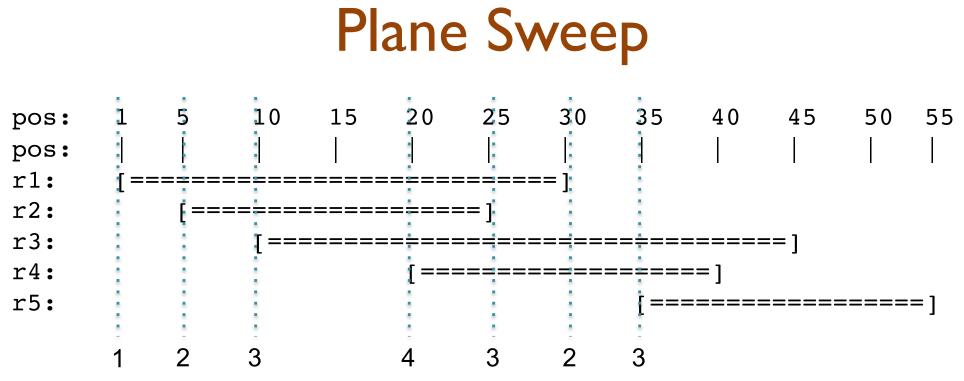


arrive at r5[35,55]:

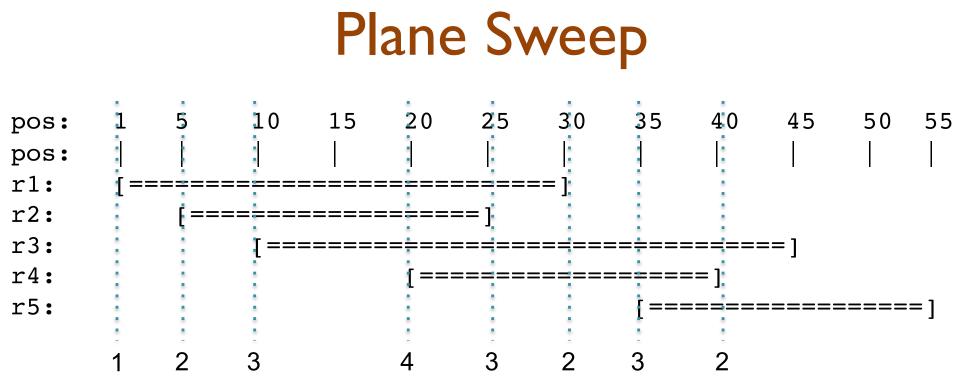
35 > 25: step down at 25; active set: 30, 40, 45 output (25, 3)

35 > 30: step down at 30; active set: 40, 45 output (30, 2)

```
35 < 40: add 55 to active set: 40, 45, 55
output (35, 3)
```

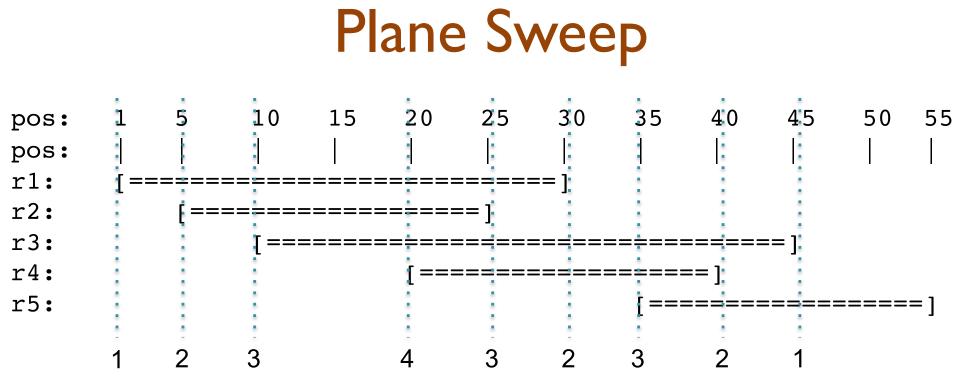


Flush: 40, 45, 55



Flush: 40, 45, 55

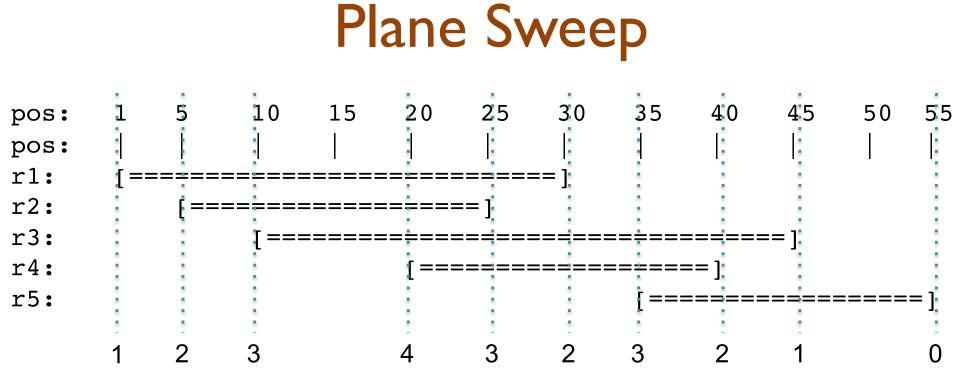
step down at 40; active set: 45, 55 output (40, 2)



Flush: 40, 45, 55

step down at 40; active set: 45, 55 output (40, 2)

step down at 45: active set: 55 output (45, 1)



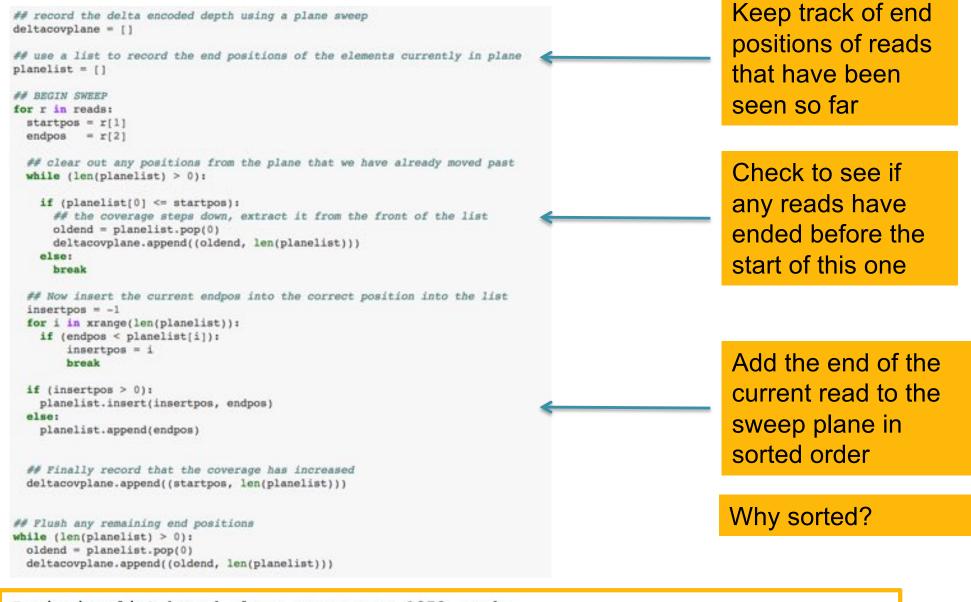
Flush: 40, 45, 55

step down at 40; active set: 45, 55 output (40, 2)

step down at 45: active set: 55 output (45, 1)

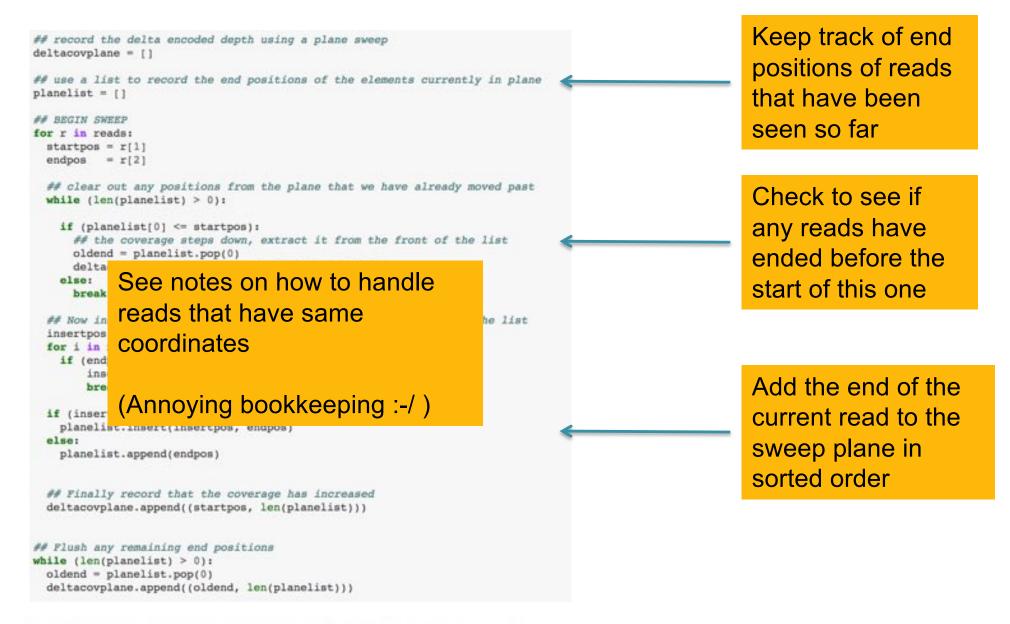
```
step down at 55: active set: {}
output (55, 0)
```

Plane Sweep



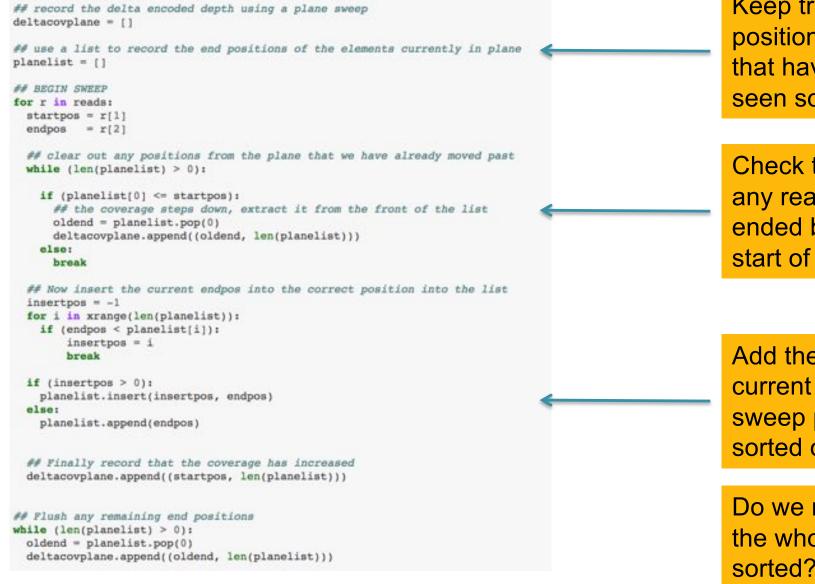
Beginning list-based plane sweep over 1873 reads Plane sweep found 3746 steps, saving 99.62% of the space in 48.90 ms (90.69 speedup)!

Plane Sweep



Beginning list-based plane sweep over 1873 reads Plane sweep found 3746 steps, saving 99.62% of the space in 48.90 ms (90.69 speedup)!

Plane Sweep



Beginning list-based plane sweep over 1873 reads Plane sweep found 3746 steps, saving 99.62% of the space in 48.90 ms (90.69 speedup)!

Keep track of end positions of reads that have been seen so far

Check to see if any reads have ended before the start of this one

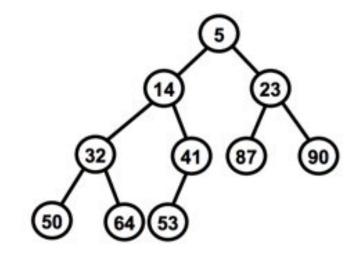
Add the end of the current read to the sweep plane in sorted order

Do we really need the whole list to be sorted?

Heaps & Priority Queues

Binary Min Heap: Binary tree such that the value of a node is less than or equal to the value of its 2 children

Similar to a binary search tree, although there are no guarantees about the relationships of the left and right children



Very efficient data structure for dynamically maintaining a set of element while allowing you to find the minimum (or maximum) very fast:

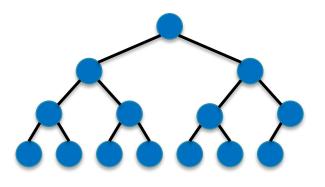
Insert: O(lg(n))	<- super fast
Remove: O(lg(n))	<- super fast
Find-min: O(1)	<- instantaneous

Key to fast performance derives from *heap shape property*: the tree is guaranteed to be a complete binary tree, meaning it will remain balanced and the height will always be log(n)

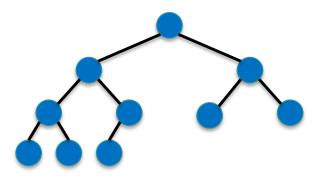
Binary Heaps

Shape Property:

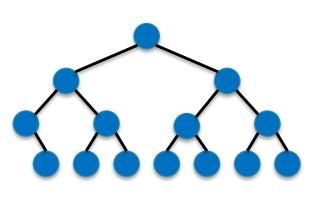
Complete binary tree with every level full, except potentially the bottom level, **AND** bottom level filled from left to right



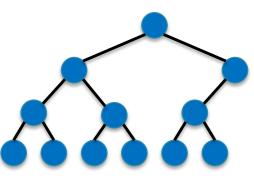
Valid



Valid



Invalid

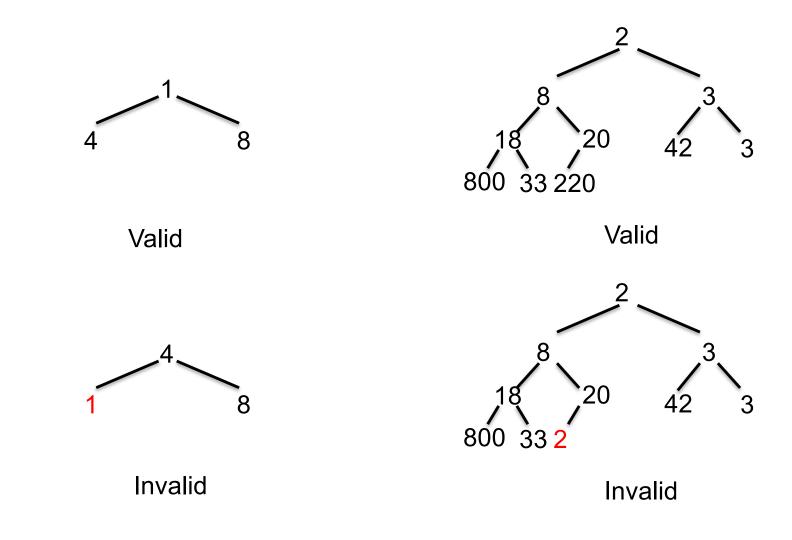


Invalid

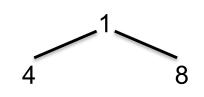
Min Binary Heaps

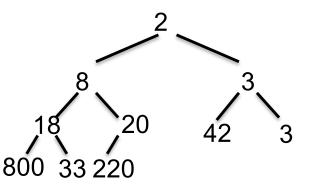
Ordering Property:

The value of each node is less than or equal to the value of its children, **BUT** there is no ordering between left and right children



Min Binary Heaps





What does the <u>shape</u> property imply about the <u>height</u> of the tree?

Guaranteed to be lg n @

What does the <u>ordering</u> property imply about the root of the tree?

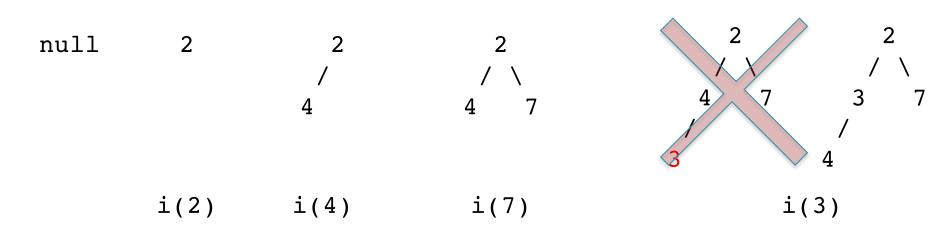
Guaranteed min (or max) value will be in the root node

That's interesting, I wonder if we could use this for a priority queue...

... just need to efficiently insert() and removeTop()

Inserting into a binary heap

Insert the elements 2, 4, 7, 3

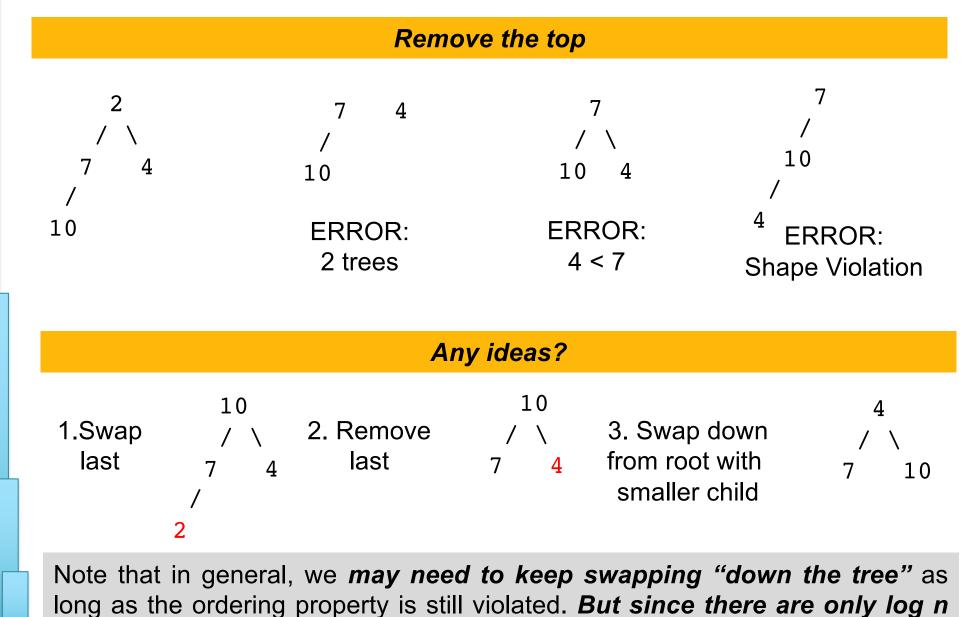


The *shape property* tells us that we need to fill one level at a time, from left to right. So the *number of elements* in a heap *uniquely determines where the next node* has to be placed.

What about the *ordering property*? When we insert 3, the parent 4 so the *ordering property is violated*. There's an *easy fix* however, just swap the values!

Note that in general, we *may need to keep swapping "up the tree"* as long as the ordering property is still violated. *But since there are only log n levels, this can take at most O(log n) time in the worst case.*

Remove top from a binary heap

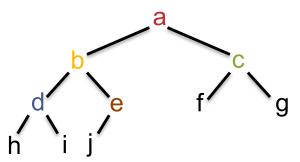


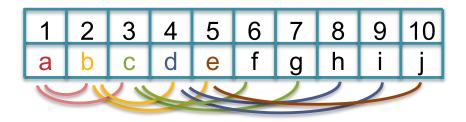
levels, this can take at most O(log n) time in the worst case.

Heap Implementation

We could implement a heap as a tree with references, but those references take up a lot of space and are relatively slow to resolve

Lets encode the tree inside an array!





Encoding a complete tree into the array in <u>level order</u> puts the children and parent in predictable locations (Math is easier if the array starts at 1 instead of 0)

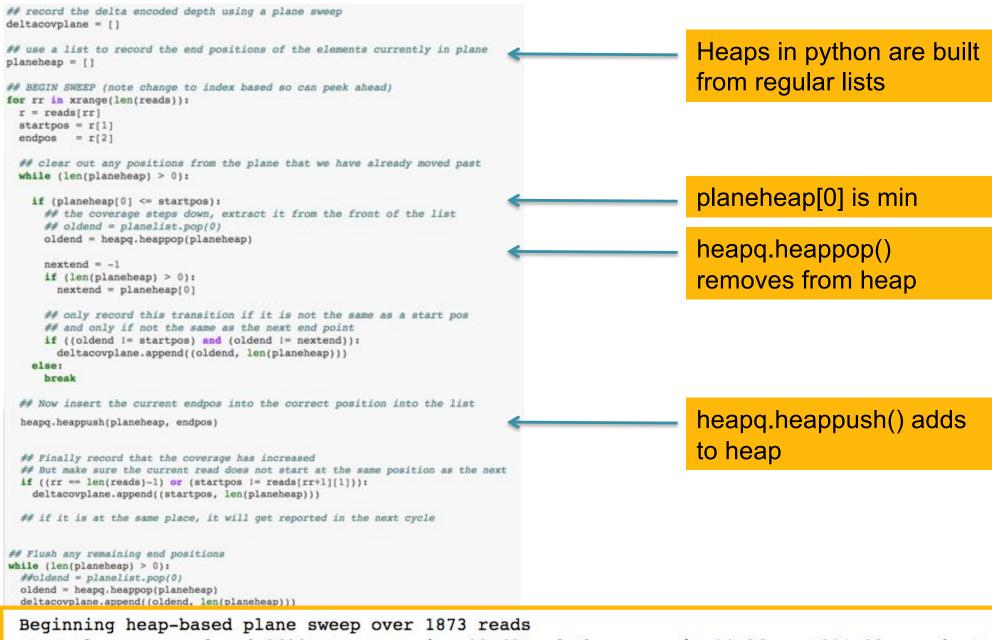
Parent(i) = array[i/2]Parent(f) = parent(6) = array[6/2] = array[3] = c

left(i) = array[i*2] & right(i) = array[i*2+1]left(3) = array[3*2] = array[6] = f & right(3) = array[3*2+1] = array[7] = g

Heaps In Python

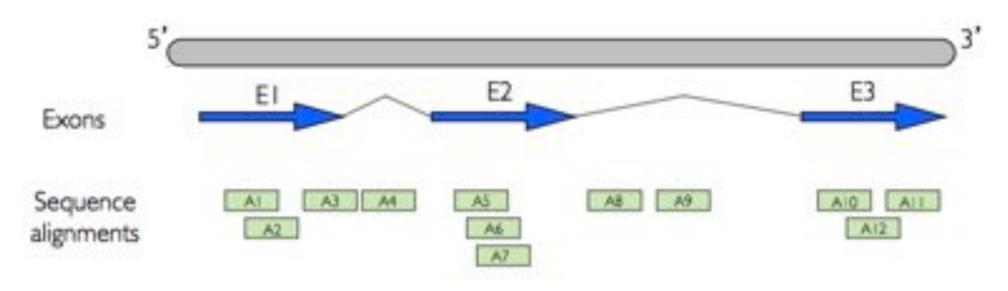
# 1 B (6 65 10015 - 10	BARRY F C	Main and a second s
and the second se	Hears Frankline (UI) Algo Hans aphon ang Unice phases, her	• • • • • • • • • • • • • • • • • • •
	II II V O Suntation O III 6 N. 12100 22 Pr El Mark 2 AM 2 Amount of Tissian 2	El Otta Basina
Printer alle Decor	nernatur - The Pyther Banded Likney - K. Dels Types -	previous 1 modules 1 inc
Table Of Contents	8.4. heapq — Heap queue algorithm	
ALL MARY - Half Coller algorithm	New in vehicle 2.3	
+ 8.4.1 Base Everyses + 8.4.9 Promy Gases	Bourse code Ut/headstay	
+ EA3 Tracy		
Previous topic	This module provides an implementation of the heap queue algorithm, also known as the priority queue algorithm.	
All exclusion in their processing and the processing elements are considered to be infinite. This implementation uses arrays for which suggests we imaginers; and an another processing elements are considered to be infinite. The interventing property of a heap is that its smallest element is always the tool, can		
Next Logic 1.5 summer - Array Summer any array	subsche ander Python uses pero-based indexing, (b) Our pop method returns the amalest term, not the largest joshed a "trim heap" in textbooks, a "max heap" is more common in texts because of to sorting.	
this Page	These bes make it possible to view the heap as a regular Python fait without surprises: surprise to the initialized fam, and surplassion maintains th	le help invariant
Peper a Reg Diver Basel	To create a heap, use a list initialized to (), or you can maneform a populated list into a heap via function immunity).	
Quick search	Ack search The following functions are provided	
	Innon, kanagpeaks/heap, Annt) Push the value dent onto the Asap, maintaining the heap invariant.	
	Pop and return the smalled laws from the heap, maintaining the heap invariant. Fifte heap is empty, <u>subsetneers</u> is raised. To access the amo	allest litem without popping it, une sequilit.
	Push item on the teap, then pip and return the smallest item from the heap. The combined action runs more efficiently than anyworky follow	well by a separate call to imaging ().
	New in version 2.6.	
	tenzes, hempi / fp()d Transform tot, a tenze, regilazze, in tenar time.	
	Pop and return the smallest item from the heap, and also push the new item. The heap size doesn't change. If the heap is empty, cubesine	is raised.
	This one slip operation is more afficient than a surgeous) followed by surgeous and can be more appropriate when using a fixed size hingings it with dom.	heep. The poplpush combination always returns an element from the heap a
	The value returned may be larger than the item added. If that an't desired, consider using suggestage;) indeed. Its push/pop continuation in	eture the smaller of the two values, leaving the larger value on the heap.
	The module also offers three-general purpose functions based on heaps.	
	Impo, anope ("Anabita) Merge multiple sorted inputs into a single sorted output (for example, marge timestamped antiries front multiple log files). Returns an lierator	over the sorted values.
	Similar to annual (second and a revealer) but returns an length, does not put the data into memory all at once, and assumes that each	h of the input ethems is already sorted (amaiwat to largest).
	New in version 2.4	. 19월 19일 : 19 19일 : 19일 : 19 19일 : 19일 : 19
	teops, #Largeen(r, downed, key)	the state of the s

Heap-based Plane-Sweep



Heap-Plane sweep found 3698 steps, saving 99.62% of the space in 14.26 ms (311.08 speedup)!

BEDTools Performance



How many reads are aligned to exonic sequences?

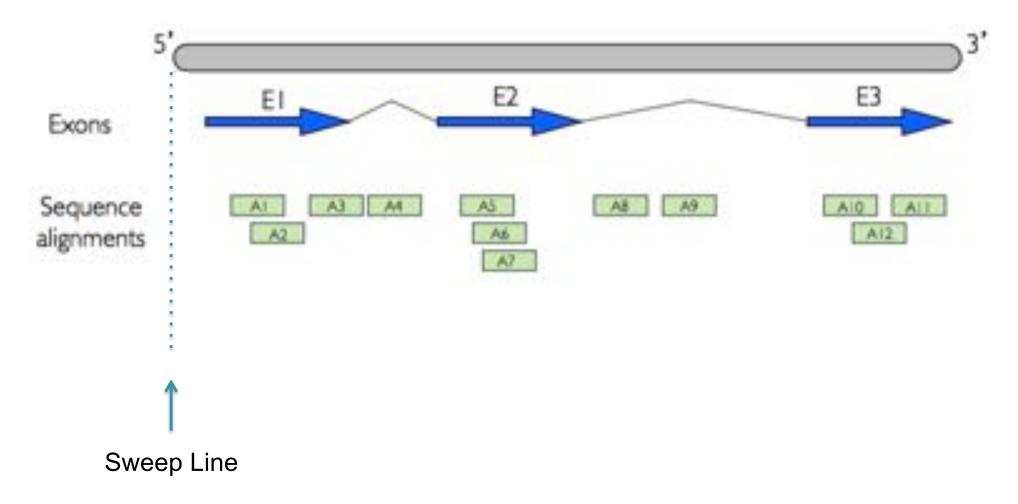
```
$ awk '{if ($3=="exon"){print}}' gencode.v21.annotation.gff3 | wc -1
1162114
```

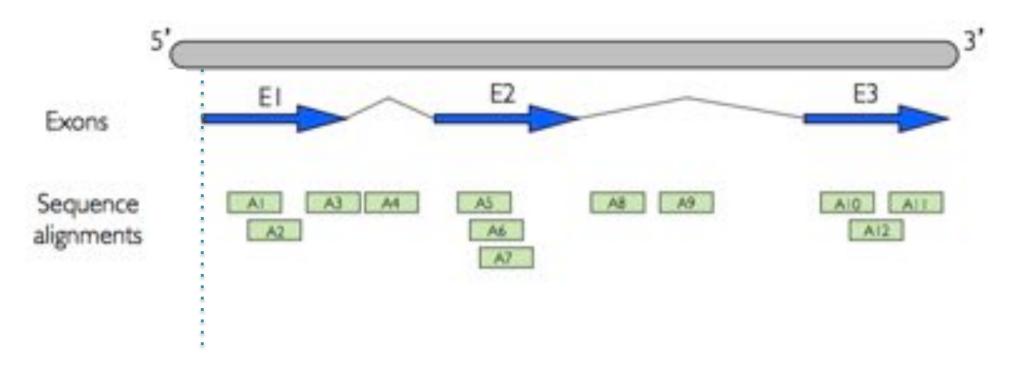
if ((read.start <= exon.end) && (read.end >= exon.start)) { print "in exon!"; }

How many comparison would a brute force approach take to scan a 30x dataset?

30x3Gb = 90Gbp / 100bp reads = 900M reads

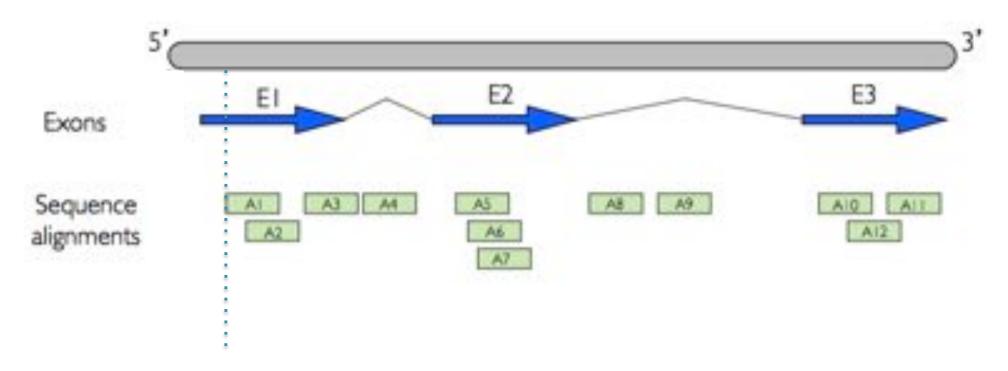
900M reads x 1.1M exons = 990MM comparisons! 😕





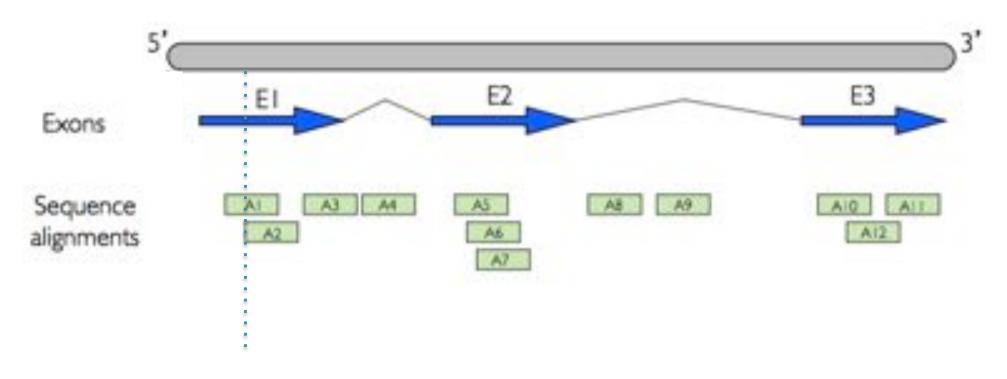
Start of E1 E1 is active

{E1}



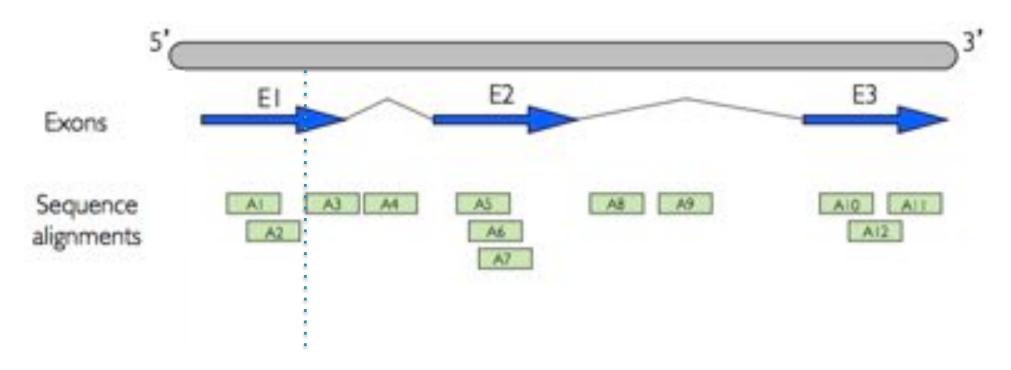
Al overlaps El

{E1=(A1)}



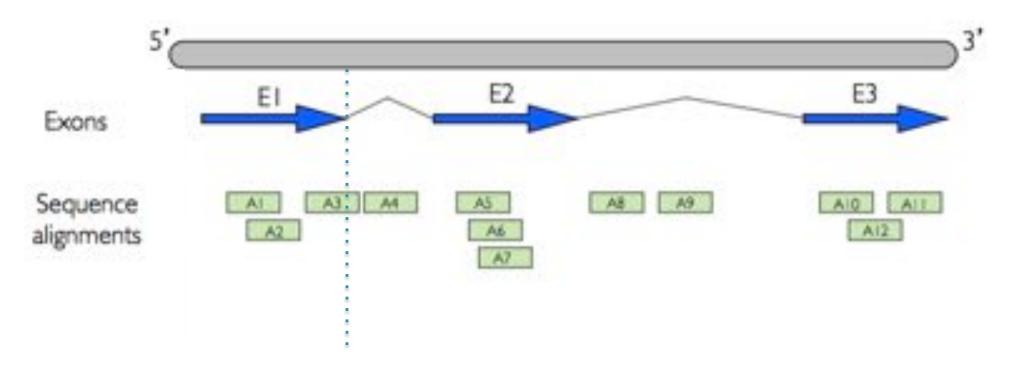
A2 overlaps E1

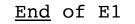
{E1=(A1, A2)}



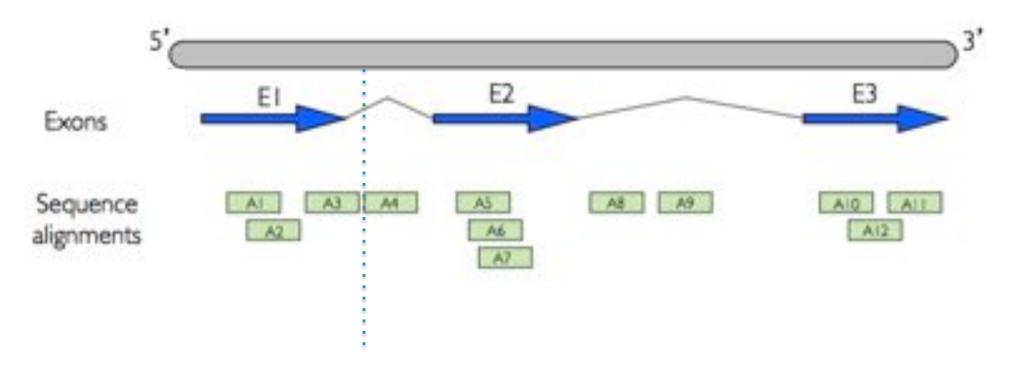
A3 overlaps E1

 $\{E1=(A1, A2, A3)\}$

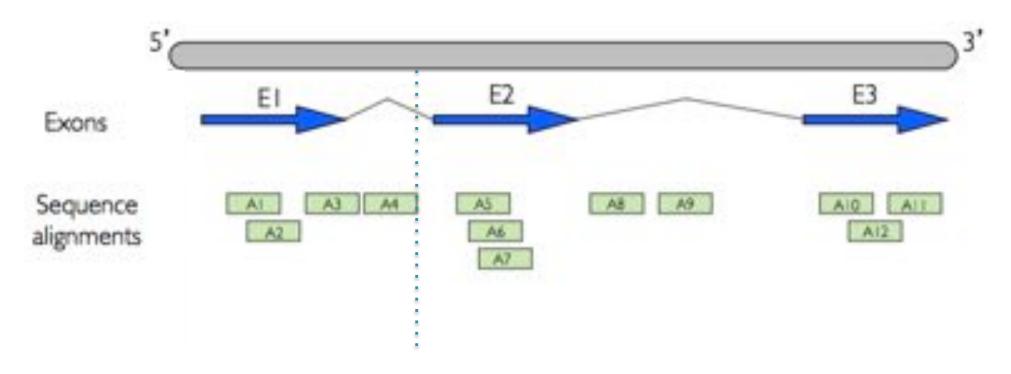




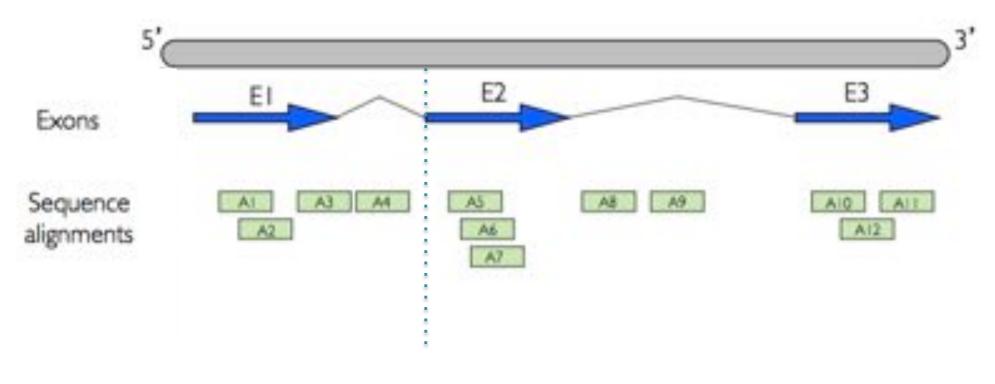
Report:
{E1=(A1, A2, A3)}



A4 starts, but nothing is active

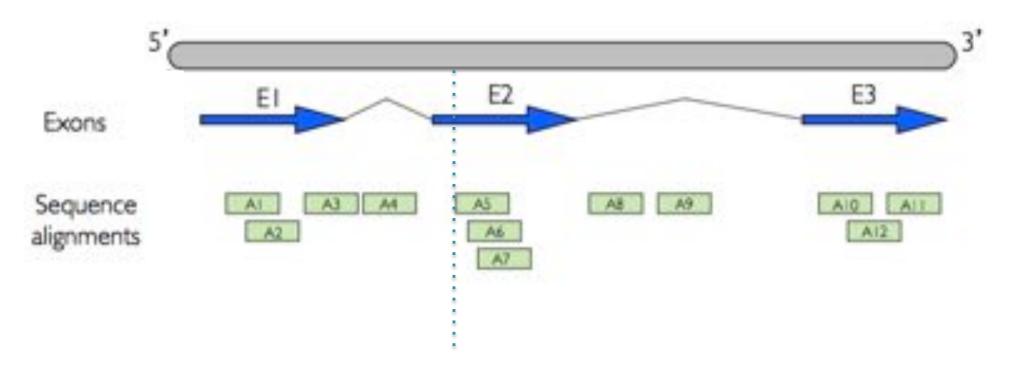


A4 end, but nothing is active



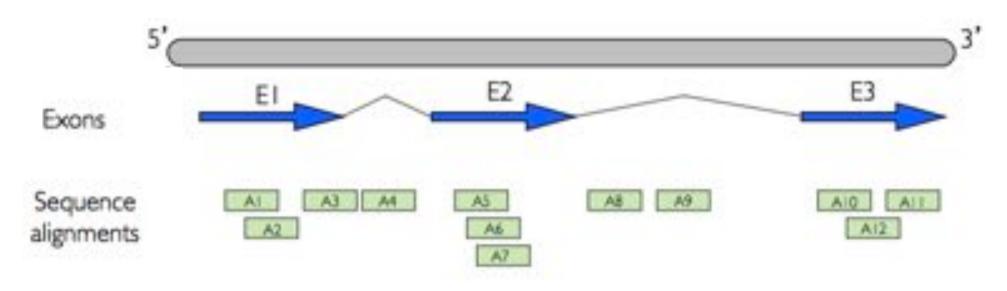
E2 starts

{E2}

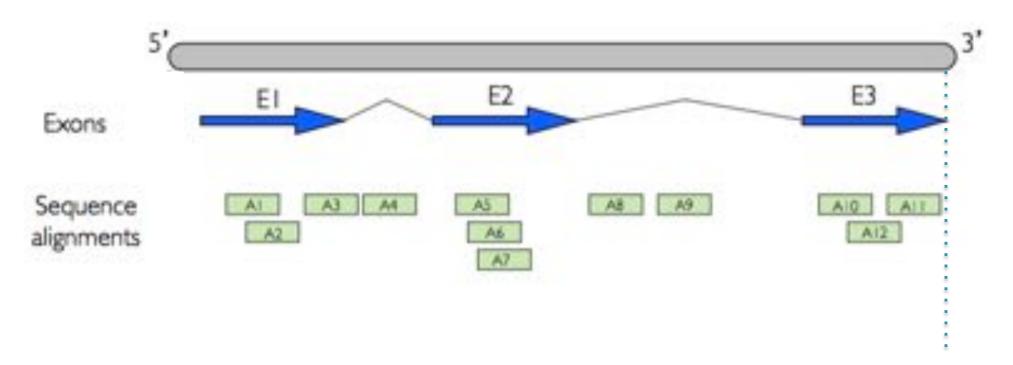


A5 overlaps E2

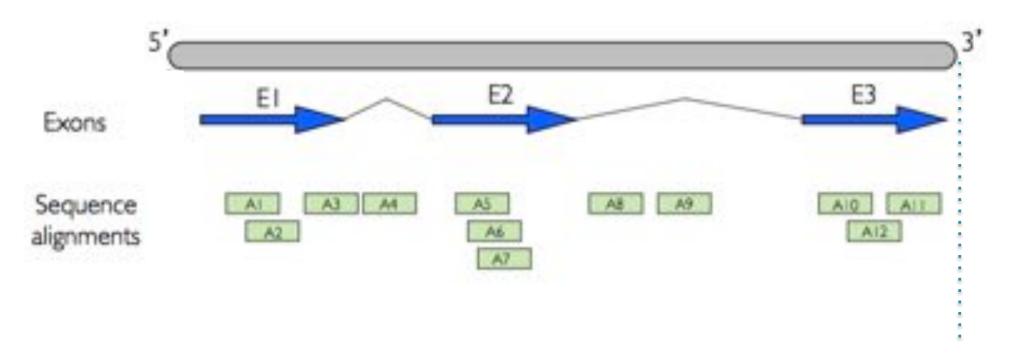
{E2=(A5)}



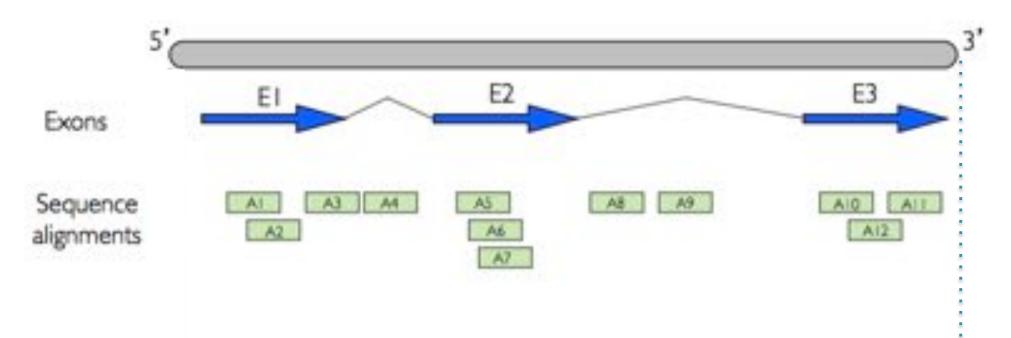
...



E3 Ends

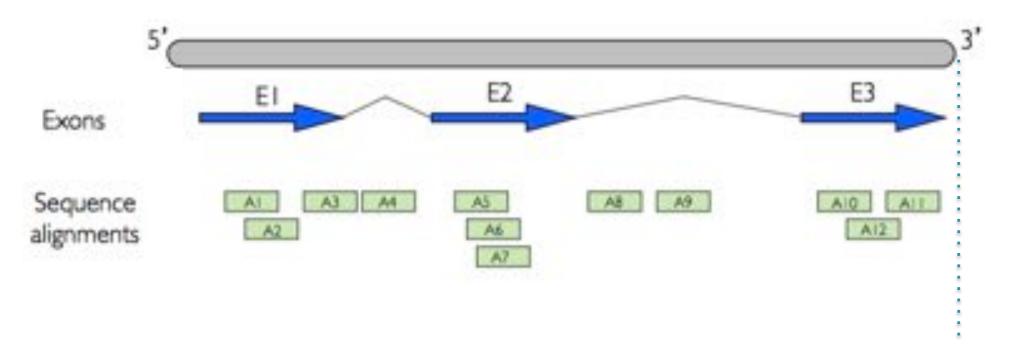


All done!



Final Results:

E1=(A1,A2,A3) E2=(A5,A6,A7) E3=(A10,A12,A11)



How many comparisons does the plane sweep algorithm make?

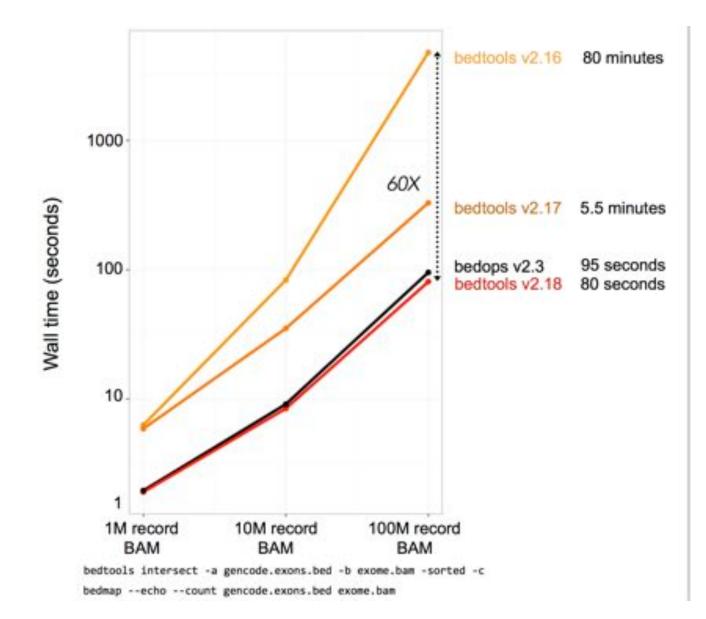
Each read is compared to the "active set"

Relatively few exons overlap: average ~1.1 active exons/position

Total comparisons: 900M reads * 1.1 "active exons/read" = 990M comparisons ©

Output is basically as fast as we can read the input data ©

BEDTools Performance



Next Steps

- I. See Lecture Notes for Full Details
- 2. Review Bedtools docs: <u>http://bedtools.readthedocs.io/</u>
- 3. Finish Assignment 3